

# Effect of Application of UV Irradiated *Beauveria bassiana* and *Metarhizium anisopliae* on Larval Weight and Mortality of *Spodoptera litura*

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## ABSTRACT

Entomopathogenic fungi have been widely used to control insect pests. The objective of this experiment was to find out the insecticidal activity of filtrate of entomopathogenic fungal cultures exposed to ultra violet (UV) C against the larvae of *Spodoptera litura*. The fungi used were *Beauveria bassiana* and *Metarhizium anisopliae* and their liquid cultures exposed to UV C (5, 10, 15, 20, and 30 watts) for 6 hours. The results showed that the larva mortality caused by *B. bassiana* culture filtrate without irradiation was the highest (97.3%) and significantly different from those caused by *M. anisopliae* culture filtrate (96.0%). However, the mortality caused by *B. bassiana* culture filtrate irradiated with UV C decreased significantly compared to the mortality caused by *M. anisopliae* culture filtrate irradiated by UV C.  $LT_{50}$  of the filtrate of *M. anisopliae* culture irradiated with UV C was 10.51 days and was significantly shorter than those of *B. bassiana* (18.09 days). Thus, the *M. anisopliae* was more resistant to irradiation compared to *B. bassiana*.

**Keywords:** *Beauveria bassiana*, insect pests,  $LT_{50}$ , *Metarhizium anisopliae*, mortality

## 1. INTRODUCTION

Freshwater swamps in Indonesia are about 9.2 Mha [1] and the land is flooded more than 6 months every year [2]. Such condition makes the soil can be planted only with specific crops that adapt to the wet conditions [3, 4]. In the dry season, farmers generally cultivate rice [5], while others cultivate several vegetables, such as cucumber, bitter-melon, yard long beans, ridge gourd [3], and chilli [6].

Chilli is a dominant vegetable crop cultivated in freshwater swamp. The main problem on chilli crop is pest attack such as *Spodoptera litura* [7], thrips [6, 8], and fruit flies [9]. An approach to reduce the population and pest attacks environmentally friendly and to produce healthy chili products is by utilizing a bio-control agents [10], i.e the use of entomopathogenic fungi. The entomopathogenic fungi have been known to be effective to various species of insect pests, such as *Beauveria bassiana* [11, 12] and *Metarhizium anisopliae* [13, 14]. *B. bassiana* culture filtrate can cause mortality up to 100% of *Spodoptera litura* larvae [11]. The effectiveness of culture filtrate is influenced by many environmental factors, hence further study needs to be conducted. The previous studies showed that light and sunlight, temperature, and humidity can affect the effectiveness of the fungi filtrate [15, 16].

Blazing sunlight can kill the fungi [17]. Short waves produced by ultraviolet (UV) radiation have been proven to reduce the viability of the entomopathogenic fungi conidia [18] and even kill the fungi [15]. The UV-B radiation at 6153.3 mW·m<sup>-2</sup> exposed for 5 minutes could decrease the germination of *B. bassiana* and *M. anisopliae* conidia from 94% to 52% and 96% to 54%, respectively [17]. Radiation of UV-B at 978 mW·m<sup>-2</sup> could cause several isolates of *B. bassiana* to be tolerant [19]. Thus, the tolerant isolates will be superior and can be developed and applied to the field. The aim of this research was to study the effect of an insecticidal activity of filtrate of entomopathogenic fungal cultures irradiated with ultra violet (UV) C on the larvae of *S. litura*.

### 1.1. Materials and Methods

This study was conducted at the Entomology Laboratory, Department of Pests and Plant Diseases, Faculty of Agriculture, Sriwijaya University from May to November 2018. The average temperature during the bioassay was 29.78°C and relative humidity was 82.72%. The isolates used in this study were explored by Safitri *et al.* (2018), namely *B. bassiana* with BSWtd2 code obtained from oil palm peat soil in Talang Dabok and *M. anisopliae*

coded MKbTp2 obtained from the highland cabbage soil in Talang Patai. Each isolate was treated with UV C irradiation with wavelengths ranged from 200-280 nm. Factorial Randomized Block Design with the first factor of 2 species of the fungi and the second factor of irradiation intensity was used. The mortality and weight data were presented in Tables 1-3.

### 1.1.1. Test insect preparation

The test insect in this experiment was *S. litura*. The larvae and eggs were collected from the synthetic pesticides-free chili crops in the experimental field of the Faculty of Agriculture, Sriwijaya University. The larvae were fed with chili leaves in a plastic cage (30 cm high x 25 cm in diameter) covered with gauze and the feed was replaced daily. When approaching the pupae stage, the last instar larvae were placed into a plastic cage containing 3 cm thick sterilized soil. Then, the pupae were transferred into an insect cage. The eggs of *S. litura* were collected by placing chilli plant. The eggs laid on the chilli leaves were transferred into the plastic cage which already provided with fresh chili leaves for feeding the newly hatched larvae. The mass rearing was carried out until getting a second generation of larvae. The third generation of the second 1-day-old instar was used as test insects in this experiment.

### 1.1.2. Preparation of the entomopathogenic fungi and production of culture filtrate

Sabouraud Dextrose Agar (SDA) medium enriched with *Tenebrio molitor* flour was used to increase fitness of *B. bassiana* and *M. anisopliae* isolates Herlinda method [10]. As many as 16.2 g of SDA medium was added with 250 ml of distilled water, then mixed with 1 g of *T. molitor* flour which had been sterilized at 100° C for 4 hours. Each culture isolate of 1 x 1 cm<sup>2</sup> of the 21 days-old SDA medium (Figure 1) was grown in SDB (Sabouraud Dextrose Broth) medium. The SDB medium was prepared in advance with as many as 30 g added 1000 ml distilled water. Then, the liquid culture (culture broth) fungi was incubated for 6 weeks (Figure 2). The culture broth for each isolate of the SDB medium was then filtered to separate the culture or supernatant filtrate from the pellets (hyphae, mycelia, and conidia/spore) through two stages using the Cheong method

(2015). A total of 100 ml culture broth on SDB was filtered into the erlenmeyer flask (500 ml volume) using Whatman No. 42 filter paper covered with 1 cm thick cotton to produce ± 70 ml of crude culture filtrate. Then, the crude filtrate culture was filtered using a syringe filter (0.45 µm-25 mm). The filtering with a syringe filter was carried out by means of 1 ml of crude culture filtrate drawn using a hypodermic needle (volume 6 ml). The needle was removed and the base of the needle was attached to a syringe filter. Then, the needle was refitted to the hypodermic needle and the 1 ml of the crude filtrate was filtered using a syringe filter to obtain culture filtrate (Figure 3).

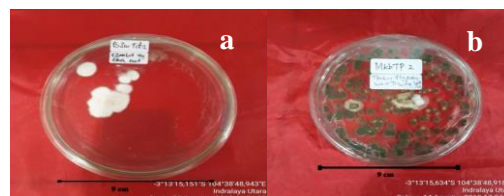
The culture filtrates were poured into Petri dish (9 cm in diameter). Then, each isolate was illuminated for 6 hours using UV C at 0, 5, 10, 15, 20, and 30 watts (= 0, 5000, 10000, 15000, 20000, and 30000 mW.m<sup>-2</sup>), and control without the fungi (distilled water). The distance between the light source and the Petri dish was 12.5 cm (Figure 4).

### 1.1.3. Bioassays for assessing insecticidal activity of the culture filtrates

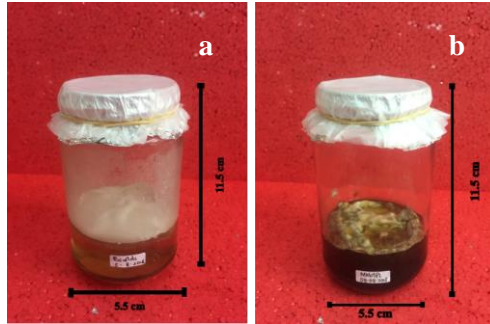
The irradiated culture filtrates were tested for their insecticidal activity against the second instar of *S. litura* larvae. As many as 5 pieces of chili leaves were dipped with the pure culture filtrate and then air-dried at room temperature. The air-dried chili leaves were put into a plastic cylinder whose top was covered with gauze (9.5 cm in diameter and 15.5 cm high), after that the 25 unfed larvae for 24 hours were introduced into the plastic cylinder. After 6 hours, the 25 larvae were transferred to another plastic cylinder containing 10 pieces of fresh leaves. Chilli leaves were replaced daily. The dead larvae were recorded and the larvae body was weighed every day for 13 days.

### 1.1.4. Data analysis

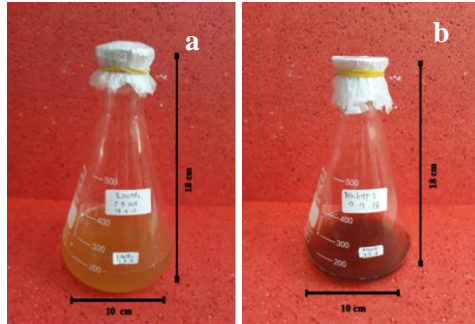
The larva mortality and weight data were analysed using analysis of variance (ANOVA) and presented in Tables 1-3. The Least Significant Difference (LSD) Test was employed to test for significant differences between treatments (isolates) at P = 0.05. LT<sub>50</sub> values were calculated by using probit analysis. All data were analysed using SAS University Edition software 2.7 9.4 M5.



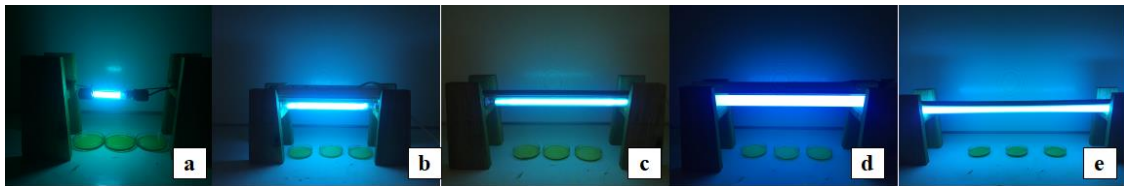
**Figure 1** Agar culture of *Beauveria bassiana* (a) and *Metarhizium anisopliae* (b) in the SDA medium



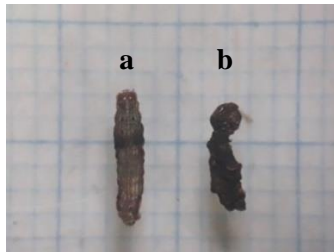
**Figure 2** Broth culture of *Beauveria bassiana* (a) and *Metarhizium anisopliae* (b) in the SDB medium



**Figure 3** Culture filtrate of *Beauveria bassiana* (a) and *Metarhizium anisopliae* (b)



**Figure 4** The radiation treatment 5 watts (a), 10 watts (b), 15 watts (c), 20 watts (d), and 30 watts (e)



**Figure 5** The healthy larvae of *Spodoptera litura* (a) and the dead one (b) caused by *Beauveria bassiana* culture filtrate (b)



**Figure 6** The healthy larvae of *Spodoptera litura* (a) and the dead one (b) caused by *Metarhizium anisopliae* culture filtrate

## 1.2. Our Contribution

This paper presents new information that the culture filtrate of *M. anisopliae* still caused the high mortality of *S. litura* larvae at the UV C irradiation of up to 30 watts for 6 hours. *M. anisopliae* could survive and was more tolerant of high irradiation intensity.

## 1.3. Paper Structure

The rest of the paper is organized as follows. Section 2 presents symptoms of *S. litura* larvae fed with the fungal culture-filtrate. Section 3 presents data of the effect of fungi on larvae weight. Section 4 shows the mortality of larvae and the time needed by 50% of dead larvae (LT<sub>50</sub>) caused by fungal culture filtrate. Finally, Section 5 concludes the paper and presents direction for future research.

## 2. RESULTS AND DISCUSSION

The *S. litura* larvae fed with the culture-filtrate treated leaves showed similar symptoms. At a day after feeding, the movement of larvae was slower than larvae fed on the untreated leaves. The feeding activity of the larvae kept declining and the body began to shrink and dull. Two days later, the larvae bodies got shrivelled, wrinkled, hard, dry, increasingly dull and black, odourless and eventually died (Figures 5 and 6). Before the larvae died, they secreted green liquid. Such larvae then grown in SDA media, after 5 to 7 days there were found no hyphae, mycelia, or conidia of the fungal. Therefore, the death of the larvae did not cause by fungi.

The data of the effect of fungi on larvae weight showed that the weight was higher on that of the *M. anisopliae* treatment than the other treatments (Table 1). However, these data were higher because they were related to the initial weights of the larvae used in the *M. anisopliae* treatment, hence the data did not reflect the influence of fungal. The data showed that the older the larvae, the higher the weight were. Yet, after 11 days of application the larva weight began to decrease.

The intensity of irradiation (UV C) to the culture filtrate of fungi was significantly affected the larvae weight 13 days after fed with the culture-filtrate treated leaves. The culture filtrate without irradiation treatment (0 watt) caused the larvae weight to drop significantly compared to the control treatment (Table 2). The culture filtrate was illuminated with irradiation intensities from 5 to 30 watt resulting in the larval weight which was not significantly different from the control using the distilled water. However, there were no significant interactions found between the fungus species and irradiation intensity.

The fungi species significantly affected the mortality of larvae and the time needed by 50% of dead larvae (LT<sub>50</sub>). The larvae mortality caused by the culture filtrate of *M. anisopliae* was significantly higher than that of *B. bassiana*. The LT<sub>50</sub> caused by the culture filtrate of *M. anisopliae*

were significantly shorter than that of *B. bassiana*. Thus, the culture filtrate of *M. anisopliae* was more effective in killing the *S. litura* larvae.

The intensity of the culture filtrate of the fungi irradiation significantly affected the mortality and LT<sub>50</sub> larvae of *S. litura*. The fungal filtrate exposed to 0 watt irradiation intensity caused the highest larva mortality (96.66%) and was significant when compared to other treatments. In addition, LT<sub>50</sub> was the shortest treatment (7.62 days) and was no significantly different from the other treatments.

Fungi species and radiation intensity significantly affected the mortality and LT<sub>50</sub> larvae of *S. litura* (Table 3). *M. anisopliae* filtrate tended to be more tolerant of the intensity of irradiation when compared to *B. bassiana*, for example when exposed to 30 watts, the larvae mortality by *M. anisopliae* was 9.33%, whereas that by *B. bassiana* was only 4%. Likewise, the LT<sub>50</sub> was affected by the species of fungi and irradiation intensity. For example, in 30-watt irradiation intensity, the LT<sub>50</sub> larvae caused by *M. anisopliae* were shorter (17.67 days) than those caused by *B. bassiana* (42.66 days).

The *S. litura* larvae feeding on leaves applied to the culture filtrates of *B. bassiana* and *M. anisopliae* showed that the symptoms of the body got shrunken, contracted, dried, and odourless. According to Ayudya et al. (2019), the insect died due to the toxic compounds contained in the culture of filtrate fungi, not due to its conidia infection. The insects died due to the conidia fungi generally showed shrivelled and hard symptoms, and from the body of the host insects grew mycelia, hyphae, and conidia fungus on the surface of the insect integument [12], whereas in this study there were no mycelia, hyphae, and conidia fungus growing in the body of *S. litura*. Consequently, the *S. litura* larvae died due to the toxic compounds contained in the culture filtrate fungus.

The intensity of the culture filtrate irradiation significantly affected the larvae weight. The irradiation intensity of 0 watt caused the larvae weight to drop significantly due to the fact that the culture filtrate did not change so that it remained effective in reducing the larvae weight. However, if the culture filtrate was illuminated with an intensity of 5 to 30 watts, the larva weight was higher than that of 0 watt intensity. This higher larval weight indicated that the culture filtrate began to decrease in effectiveness. The culture filtrate was less able to reduce appetite of the larvae and they remained healthy with normal weight like those of treated with distilled water (control).

Although the irradiation of 5 to 30 watts against the culture filtrate of the fungi began to decrease the effectiveness of the fungi, the *M. anisopliae* was more tolerant of the UV C irradiation than *B. bassiana*. It is new information that the culture filtrate of *M. anisopliae* still caused the high mortality of *S. litura* larvae despite the UV C irradiation of up to 30 watts for 6 hours; the mortality still reached 9.33%. *M. anisopliae* could survive and was more tolerant of high irradiation intensity because this fungus had a darker pigment which was more resistant to UV light than

of the white fungus such as *B. bassiana* [20]. However, in the application of the entomopathogenic fungi in the field, it is still necessary to consider that the application should be carried out in the morning or evening when the sun does not

shine brightly. The main problem in the utilization of the fungus is because of the low tolerance to sunlight.

**Table 1** The effect of species fungi on larvae weight of *Spodoptera litura* larvae

Species of fungi	Larvae weight (g/larvae) after fungal filtrate UV treatment						
	1 day	3 days	5 days	7 days	9 days	11 days	13 days
<i>Beauveria bassiana</i>	0.30 <sup>a</sup>	0.58	1.08 <sup>a</sup>	1.64 <sup>a</sup>	4.42	3.14	3.01
<i>Metarhizium anisopliae</i>	0.38 <sup>b</sup>	0.59	1.19 <sup>b</sup>	1.81 <sup>b</sup>	4.31	3.28	3.53
ANOVA F-value	34.56*	0.27 <sup>ns</sup>	8.75*	5.61*	0.52 <sup>ns</sup>	0.28 <sup>ns</sup>	2.69 <sup>ns</sup>
P value (0.05)	0.001	0.61	0.01	0.02	0.47	0.60	0.11
LSD test	0.01	-	0.02	0.03	-	-	-

Note: \* = significantly different; values within a column followed by the same letters were not significantly different at  $P < 0.05$  according to LSD test, ns = not significantly different

**Table 2** The effect of the intensity of UV irradiation for fungal filtrate on larvae weight of *Spodoptera litura* larvae

Intensity of UV irradiation	Larvae weight (g/larvae) after UV fungal filtrate treatment						
	1 day	3 days	5 days	7 days	9 days	11 days	13 days
Control (aquadest)	0.33	0.61	1.11	1.66	4.51	3.24	3.59 <sup>ab</sup>
0 watt	0.34	0.54	1.08	1.70	4.76	3.34	1.39 <sup>a</sup>
5 watts	0.33	0.56	1.24	1.78	4.58	3.36	3.38 <sup>ab</sup>
10 watts	0.33	0.57	1.13	1.66	3.92	3.08	3.37 <sup>ab</sup>
15 watts	0.36	0.55	1.18	1.83	4.40	3.18	3.82 <sup>ab</sup>
20 watts	0.35	0.64	1.10	1.72	4.23	2.92	3.75 <sup>ab</sup>
30 watts	0.36	0.62	1.11	1.72	4.16	3.31	3.60 <sup>ab</sup>
ANOVA F-value	0.58 <sup>ns</sup>	0.78 <sup>ns</sup>	1.44 <sup>ns</sup>	0.42 <sup>ns</sup>	0.61 <sup>ns</sup>	0.20 <sup>ns</sup>	4.07*
P value (0.05)	0.74	0.59	0.23	0.853	0.72	0.973	0.01
LSD test	-	-	-	-	-	-	0.49

Note: \* = significantly different; values within a column followed by the same letters were not significantly different at  $P < 0.05$  according to LSD test, ns = not significantly different

**Table 3** The effect of fungal species and the intensity of UV irradiation for fungal filtrate on mortality and  $LT_{50}$  of *Spodoptera litura* larvae

Species of fungi x intensity of irradiation	Mortality (%)	$LT_{50}$ (days)
Control (aquadest)	0 <sup>a</sup>	-
<i>B. bassiana</i> x 0 watts	97.33±2.67 <sup>hi</sup>	6.41±0.24 <sup>ab</sup>
<i>B. bassiana</i> x 5 watts	54.66±1.33 <sup>f</sup>	12.16±0.31 <sup>bc</sup>
<i>B. bassiana</i> x 10 watts	32.00±2.31 <sup>d</sup>	16.95±1.50 <sup>bcd</sup>
<i>B. bassiana</i> x 15 watts	30.66±4.81 <sup>d</sup>	20.59±3.35 <sup>cd</sup>
<i>B. bassiana</i> x 20 watts	12.00±2.31 <sup>c</sup>	27.85±16.37 <sup>d</sup>

Table 3 (continuation)

<i>B. bassiana</i> x 30 watts	4.00±2.31 <sup>b</sup>	42.66±29.80 <sup>e</sup>
Control (aquadest)	0 <sup>a</sup>	-
<i>M. anisopliae</i> x 0 watts	96.66±4.00 <sup>h</sup>	7.62±0.20 <sup>ab</sup>
<i>M. anisopliae</i> x 5 watts	68.00±4.00 <sup>g</sup>	9.24±0.34 <sup>abc</sup>
<i>M. anisopliae</i> x 10 watts	50.67±35.3 <sup>ef</sup>	10.98±0.32 <sup>bc</sup>
<i>M. anisopliae</i> x 15 watts	38.66±5.81 <sup>de</sup>	12.51±1.68 <sup>bc</sup>
<i>M. anisopliae</i> x 20 watts	26.60±2.67 <sup>d</sup>	15.58±1.40 <sup>bc</sup>
<i>M. anisopliae</i> x 30 watts	9.33±2.67 <sup>c</sup>	17.67±1.71 <sup>bcd</sup>
ANOVA F-value	1.248*	4.300*
P value (0.05)	0.00	0.00
LSD test	5.25	11.56

Note: \* = significantly different; values within a column followed by the same letters were not significantly different at P < 0.05 according to LSD test, ns = not significantly different

### 3. CONCLUSION

*Metarhizium anisopliae* is more tolerant of irradiation compared to *B. bassiana*. This implies that *M. anisopliae* has more potential to survive in agroecosystems with relatively more intense sunlight such as in the tropical lowlands such as freshwater swamps.

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