

# Feed Intake and Feed Digesbility of Male Merino Sheep with Adding Mahagany Leaves (*Swietania mahagoni*) as a Source of Tannins in Feed

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#### ABSTRACT

The effect of adding Mahogany leaves as a source of tannins in feed on nutrient intake and digestibility in merino sheep was determined in this study. This study used 12 male sheep with an average body weight of 30 to 35 kg. They were confined in individual cages and given feed two times a day. To separate feces and urine samples, there were nylon nets and a urine reservoir under the cages. The feed was given treatments of tannins addition of 0%, 1.5%, and 3%. The 14-days adaptation period and 7-days collection period were conducted to obtain samples: forages feed samples, concentrate, forage residue, and stool. These samples tested for their nutrient composition, such as dry matter (DM), organic matter (OM), crude protein (CP), extract ether (EE), crude fiber (CF), and extract without nitrogen (ETN) content. The data obtained were used to determine the value of nutrient intake and digestibility. The result showed that nutrient intake was decreased at added tannins at 1.5% and 3% (P <0.05). In contrast, the addition of 3% tannins increased nutrient digestibility. It is concluded that Mahogany leaves could be added in merino feed with a level of tannins of 3%.

Keywords: Digestibility, Mahogany leaves, nutrient intake, sheep, tannins.

## 1. INTRODUCTION

Merino sheep are one of the famously developed ruminants in Indonesia. However, one of the obstacles that usually occur in raising merino sheep is the fulfillment of nutrients, especially the availability of protein that escapes rumen degradation. Ruminants have a rumen in which the degradation of protein from feed into microbial protein happens then the products are used as the primary source of protein. Still, protein sources that come from microbial protein cannot engage the protein needs of livestock [1]. Thus, adding a higher protein concentrate helps to fulfill protein requirements for livestock [2]. A protein protection agent mostly accompanies the addition of protein concentrate. It aims to protect the proteins from rumen microbes degradation. It is known that tannins are protein protection agents capable of binding proteins. Tannins and proteins will form a tannin-protein complex bond that will escape rumen microbial degradation. The complex bonds are digested in the abomasum and intestines since these bonds will be broken down in an acidic and alkaline environment [3]. As a legume, Mahogany contains tannins in its leaves. The addition of Mahogany leaves in a certain amount can protect the protein in the feed [4].

Based on the explanation above, it is necessary to evaluate nutrient intake, and digestibility from the male Merino sheep feed ratio with Mahogany leaves as a protein protection agent.

#### 2. MATERIALS AND METHODS

#### 2.1 Animals and Feeding Treatment

The livestock was fed with concentrate and king grass. The drinking water and feed were given ad libitum by giving the ratio of 3% of the dry matter (DM) requirement per body weight. When the feed and drinking water were fully consumed, they were given extra 10% addition of the first feed requirement. Mahogany leaves as a source of tannins were dried and ground. The treatment of this research was the addition level of Mahogany leaves by 0%, 1.5%, and 3% based on dry matter of feed.

Feed ingredient	DM (%)	OM (%)	CP (%)	EE (%)	CF (%)	ETN (%)	Tannins
King grass	17.96	86.90	6.85	0.31	32.26	48.20	-
Concentrate	89.32	88.33	15.18	2.62	18.06	52.48	-
Mahogany leaves	88.59	88.16	12.61	1.74	26.56	47.25	10.62

Table 1. Feed chemical composition

# 2.2 Collection period

All livestock were weighed before carrying out the study to determine the initial body weight. The sheep were caged in the individual metabolic pen, while underneath it is given a stool box and a urine reservoir to separate feces and urine. There was an adaptation period of 14 days and then continued with a collection period of 7 days. There were 12 merino sheep used, 10 to 12 months, with an average body weight of 30 to 35 kg. The samples collected during the collection period were forage, concentrate, feed residue, and feces. The remaining feed and feces were weighed and taken 10% as samples and stored at 5°C for 2 days. Each sample was weighed and mixed to get subsamples for chemical analysis.

## 2.3 Chemical analysis

Forage and feed residue was ground on a 1 mm screen for chemical analysis, while the feces samples were analyzed in fresh condition. Each sample was analyzed with proximate analysis of dry matter (DM), organic matter (BO), crude protein (CP), extract ether (EE), crude fiber (CF) based on the AOAC method [5]. Extract without nitrogen (ETN) was calculated from the results of this analysis. Meanwhile, nutrient intake and digestibility were calculated based on the results of the chemical analysis of feed and feces.

## 2.4 Statistical Analysis

The results of the research data were analyzed using one-way ANOVA. Duncan's New Multiple Range Test (DMRT) was done to show different significance [6].

## **3. RESULTS AND DISCUSSION**

This experiment fed Merino sheep, including king grass, concentrate, and tannins from Mahogany leaves. Table 1 presents the chemical composition of each feed ingredient. According to the analysis result, it is known that Mahogany leaves contain 10.62% of total tannins. *Swietenia mahagoni* has total tannins of as much as 11.92% and can bind protein higher than *Leucaena leucocephala* [7]. The feed consisted of forage and concentrated with the ratio 60:40 (w/w DM basis). The content of CP and TDN in the control feed was 10.18% and 47.26%. However, increasing the feed CP by 0.04-

Table 2. Nutrient intake of Merino sheep with different levels of tannins addition

Parameters	Tannins level (%) DM basic					
	0	1.5	3			
In g/day						
DM	986.80±59.33 <sup>c</sup>	749.65±30.03ª	915.30±33.30 <sup>b</sup>			
OM	861.82 ±51.40°	650.79±27.61ª	798.02±31.27 <sup>b</sup>			
СР	102.77±6.73 <sup>b</sup>	85.55±5.04ª	98.78±3.18 <sup>b</sup>			
EE	12.05±0.83 <sup>b</sup>	9.53±0.43ª	11.58±0.62 <sup>b</sup>			
CF	252.43±14.29 <sup>c</sup>	180.35±8.75ª	225.36±13.14 <sup>b</sup>			
ETN	494.57±29.71°	375.35±16.38ª	460.95±12.94 <sup>b</sup>			
In g/(d kg BW <sup>0.75</sup> )						
DM	71.83±0.57 <sup>c</sup>	56.92±3.99 <sup>a</sup>	66.14±3.73 <sup>b</sup>			
OM	62.74±0.47 <sup>c</sup>	49.43± 3.52ª	57.68±3.30 <sup>b</sup>			
СР	7.47±0.81 <sup>b</sup>	6.50±0.29ª	7.21±0.27 <sup>b</sup>			
EE	0.89±0.02 <sup>b</sup>	0.73±0.03ª	0.84±0.06 <sup>b</sup>			
CF	18.39±0.13 <sup>c</sup>	13.69±1.18ª	16.33±1.18 <sup>b</sup>			
ETN	36.00±0.23°	28.51±2.02ª	33.29±1.76 <sup>b</sup>			

<sup>abc</sup> Different superscript at the same column shows significantly different p < 0.05)

Development	Tannins level (%) DM basic						
Parameters	0	1.5	3				
In g/day							
DM	658.96±46.42 <sup>c</sup>	459.16±42.19 <sup>a</sup>	594.93±20.85 <sup>b</sup>				
OM	606.28±40.68°	429.36±33.9ª	549.45±22.16 <sup>b</sup>				
СР	62.76±4.19 <sup>b</sup>	51.97±5.23ª	62.66±4.27 b				
EE	5.52±0.48 <sup>b</sup>	4.26±0.31ª	5.67±0.25 b				
CF	157.86±5.02 <sup>b</sup>	107.03±10.86ª	152.11±20.07 <sup>b</sup>				
ETN	380.12±31.52 <sup>b</sup>	266.10±21.01 <sup>a</sup>	342.98±5.89 <sup>b</sup>				
In g/(d.kg BW <sup>0.75</sup> )							
DM	47.95±0.95°	34.58±3.20 <sup>a</sup>	43.06±2.19 <sup>b</sup>				
OM	44.12±0.78 <sup>c</sup>	32.39±2.91ª	39.75±1.99 <sup>b</sup>				
СР	4.57±0.03 <sup>b</sup>	3.93±0.23 <sup>a</sup>	4.50±0.14 <sup>b</sup>				
EE	0.41±0.12 <sup>b</sup>	0.33±0.01ª	0.40±0.02 <sup>b</sup>				
CF	11.50±0.28°	8.15±0.29ª	10.01±0.62 <sup>b</sup>				
ETN	27.65±0.65 <sup>c</sup>	19.98±1.73ª	24.82±0.54 <sup>b</sup>				
In %							
DM	66.75±1.67 <sup>b</sup>	60.45±1.36 <sup>a</sup>	65.09±0.93 <sup>b</sup>				
OM	70.32±1.53 <sup>b</sup>	65.27±1.27ª	68.94±0.61 <sup>b</sup>				
СР	61.18±0.62 <sup>ab</sup>	60.48±0.56ª	62.48±1.52 <sup>b</sup>				
EE	45.71±1.59ª	44.96±2.07ª	49.78±0.84 <sup>b</sup>				
CF	62.52±1.01 <sup>b</sup>	59.61±1.59ª	60.75±1.27 <sup>ab</sup>				
ETN	76.81±0.11 <sup>b</sup>	69.60±3.67ª	74.73±1.64 <sup>b</sup>				

Table 3. Nutrient digestibility of Merino sheep with different levels of tannins addition

<sup>a, b, ab, c</sup> Different superscript at the same column shows significantly different (P <0.05)

0.05 % and TDN by 0.14-0.23 % by adding Mahogany leaves as a tannins source by 1.5 and 3 % only increases the feed CP by 0.04-0.05 % and TDN by 0.14-0.23 %.

## 3.1 Nutrient intake of Merino sheep

The results in Table 2 show a significant difference (P <0.05) in the nutrient consumption of Merino sheep with tannins addition of 0%, 1.5%, and 3% based on DM of feed. All nutrient intake with the addition of Mahogany leaves as tannins source were decreased compared to control feed, yet still, fulfill the livestock requirement for daily maintenance. Nutrient consumption decreased at tannins addition of 1.5% but increased at 3%. Dry matter consumption showed an average value of 2.7% body weight, higher than the necessity of life, namely 2.5% - 2.7% for bodyweight 30-40 kg [8]. Sheep with a bodyweight of 50 kg performed 2% DM intake from body weight and CP intake as much as 95 g/head/day [9]. Meanwhile, sheep with a bodyweight of 30 up to 40 kg had 62 - 77 g/head/day of CP intake [8]. Nutrient consumption in this study was not much different from the nutrient consumption of previous studies on male merino sheep fed with peanut straw as sole feed [10]. Animal weight and type of feed can affect nutrient consumption, where the higher the body weight, the greater the nutrient needs to meet their daily needs [11,12]. However, the addition of tannins to feed can affect palatability in livestock, thereby reducing feed consumption [13].

# 3.2 Nutrient Digestibility

Table 2 shows nutrient consumption in Merino sheep with a significant result (P <0.05). Moreover, the nutrient digestibility results (Table 3) showed that higher levels of tannins showed a significant increase (P<0.05) of digestibility values. According to the result, the addition of 3% Mahogany leaves increased DM, OM, CF, and ETN 10%, also EE 22% compared to control. Protein digestibility with tannins treatments 1.5% and 3% showed higher yields 3% and 1.5% compared to control, respectively. High protein digestibility indicates the fast rate of emptying the contents of the rumen. Due to



tannins that bind protein, the feed escapes and is digested post-rumen [14]. The condensed tannins present in animal feed causes low levels of protein digestibility in the rumen [15]. Increased protein consumption is closely related to the digestibility of feed protein [16]. Tannins and protein bonds are influenced by pH [17], where the tannins-protein complex will be broken down in lower pH conditions (pH<7) in the abomasum [18]. The characteristic of the tannins bond leads to protein protection (by-pass) from rumen microbial degradation, hence increasing the hosts' protein availability.

#### **4. CONCLUSION**

In conclusion, adding tannins up to 3% increases the value of nutrient digestibility, allowing for more efficient feed nutrient utilization.

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## REFERENCES

- [1] C.S. Ali, I.U. Din, M. Sharif, M. Nisa, A. Javaid, N. Hasmi and M. Sarwar.2009 Supplementation of ruminally protected proteins and amino acids: feeds consumption, digestion and performance of cattle and sheep. Int. J. Agric Biol, vol. 11, 2009, pp. 477-482.
- [2] H. Hartadi, Feed Composition Table for Indonesia. Gajahmada University Press, 2005.
- [3] R.D. Cahyani, LK Nuswantoro., And A. Subrata, Effect of protection of soy flour with mangrove leaf tannins on the concentration of ammonia, undegraded protein and total protein in vitro, Animal Agricultur journal, vol 1,2012. Pp 159-166.
- [4] T. Robinson, High Plant Organic Content (Translation) Padmawinata, K, ITB, 1995.
- [5] AOAC. Official Method of Analysis of the Association of Official Analytical Chemists 18 th ed, Maryland: AOAC International, ed William Harwitz United States of America, 2005.
- [6] R.D.G. Steel, and JH Torrie. Principles and Procedures of Statistics 2<sup>nd</sup> ed New York: McGraw-Hill Book Company, 1980.
- [7] L. M., Yusiati. A. Kurniawati, C. Hanim, and M. A. Anas. Protein binding capacity of different forages tannin. In: IOP Conf. Series: Earth and Environmental Science, vol 119, Semarang 2018,

Pp 1-5. https://doi:10.1088/1755-1315/119/1/012007

- [8] NRC, Nutrient requirements of goats: angora, dairy, and meat goats in temperate and tropical countries Nutrient Requirements of Domestic Animal (Washington DC: National Academy Press, 1981.
- [9] P.R. Cheeke, Applied Animal Nutrition Feeds and Feeding. 3rd Ed. Pearson Prentice Hall, 2005.
- [10] C. Hanim, and Muhlisin. Nutrient intake and digestibility in Merino sheep fed peanut straw. IOP Conf. Series: Earth and Environmental Science Vol 119 Semarang, 2018.Pp 1-5. https://doi:10.1088/1755-1315/119/1/012005
- [11] N. Ngadiyono., E. Baliarti., T.S.M. Widi., H. Maulana and B.A. Atmoko. Effect of breed and initial body weight on daily weight gain of Simmental Ongole Crossbred cattle and Ongole Grad cattle. IOP Conf. Series: Earth and Environmental Science Vol 387, Yogyakarta 2019, Pp 1-4. https://doi:10.1088/1755-1315/387/1/012044
- [12] J.M. Wilkinson, and B.A Stark, Commercial Goat Production. 1 st Ed. BSP Professional Books, 1987.
- [13] E. Haslam, Vegetable tannins. In Biochemistry of Plant Phenolics editors: T Swain, JB Harborne and C Van Sumere). The Plenum of London, 1979.
- [14] T.J Kempton, J.V. Nolan, and R.A. Leng. 1977. Principle for the use of non-protein nitrogen and by-pass protein in diets of ruminants. World Animal Review, vol 22, 1997 pp.160-181.
- [15] J.J. Karchesy, and R.W. Hemingway. 1986; Condensed tannins:  $(4\beta \rightarrow 8; 2\beta \rightarrow O \rightarrow 7)$ -linked procyanidins in Arachis hypogea, LJ Agric. Food Chem. Vol 34, 1986 pp. 966-970.
- [16] A.D. Tillman, H. Hartadi, S. Reksohadiprodjo, S. Prawirakusomo and S. Lebdosoekojo, Basic Food Science. Gadjah Mada University Press, 1991.
- [17] M. Naczk, D. Oickle, D. Pink and F. Shahidi, Protein precipitating capacity of crude canola tannins: effect of pH, tannin, and protein concentrations. Journal of Agricultural and Food Chemistry, vol 44, 1996 pp. 2144-2148. https://doi.org/10.1021/jf960165k
- [18] Diaz-Hernandez, A., M.J. Nixon, F.M. Ball, R.A. Leng, and J.B. Rowe, Protein-tannin complexes are susceptible to proteolytic degradation. Proc. of The New Zealand Society of Animal Production. Vol 57, 1997, 116-119.