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# A Review and Current Research on Biomaterials Supporting Artificial Insemination Technology Advancement

Enike Dwi Kusumawati<sup>1\*</sup>, I Wayan Karyasa<sup>2</sup>

<sup>1</sup> Faculty of Animal Husbandry, Universitas PGRI Kanjuruhan Malang

<sup>2</sup> Chemistry Department, FMIPA, Universitas Pendidikan Ganesha

\*Corresponding author. Email: <u>enike@unikama.ac.id</u>

#### ABSTRACT

ATLANTIS

The development of artificial insemination (AI) technology today plays an important role in the animal husbandry industry. The technology involves the enhancement of AI processes as well as improvement related to supporting biomaterials. A brief systematic literature review was aimed at proposed research problems in order to make the current development of artificial insemination becomes more effective and efficient as well as productive in securing our sustainable food. The study found that AI has still a lack in effectiveness and cost because of cryopreservation needs freezing step using liquid nitrogen. We proposed that developing better techniques and biomaterials supporting AI, can be not only low cost but also eco-friendly; thus, they contribute to the sustainable development of food for all people leading to the human beings as well as animal welfare.

Keywords: artificial insemination, biomaterial, AI process.

# **1. INTRODUCTION**

One of the major developments of animal breeding is artificial insemination (AI) because of its important role in genetic improvement, genetic resource conservation, and productivity in animal husbandry. AI technology is a part of reproductive technologies besides multiple ovulation and embryo transfer (MOET), and in vitro embryo production (IVEP), which can lead to genetic improvement of animal husbandry and also has a major role to conserve animal genetic resources for all of the major domesticated species [1]. As an effective assisted reproductive technology, AI can also ensure global food security through sustaining the productivity of ruminant livestock because of its ability to up-cycle non-human-edible products into meat and milk products with notable nutritional value [2]. Hence, the advancement of AI technology is a strategic way for our global food security.

The development of AI technology involves the enhancement of AI processes as well as improvement related to supporting biomaterials. The sustainable improvement of AI processes and their supporting biomaterials is purposed to ensure the effectiveness and efficiency of assisted reproductive animal breeding. In order to reach the purpose, particularly, it is a need for researches on the biomaterials supporting AI technology. Our paper reported a systematic literature review results and current laboratory works in that area.

## 2. MATERIALS AND METHODS

A systematic literature review (SLR) [3] was conducted to provide an accurate and comprehensive unbiased synthesis of relevant searched literatures regarding biomaterials supporting processes of artificial insemination technology.

We searched electronic databases using search engines, namely Google Scholar, Research Gate, Garuda, and Scopus for gathering the published last decade (2017-2021) articles (research paper, review, thesis, dissertation and conference proceeding as well as patents) in September 2021. We adopted the PICOS [4] (P = problem, I = intervention or solution, C = comparators, O = outcomes, S = study type) format for formulating research questions focusing on materials and processes involving the steps in performing artificial insemination technology for husbandry animal reproduction. The search was performed with keywords combined with Boolean operators AND, OR, and NOT.

There were totally 18,000 publications retrieved from the databases by using key words artificial insemination. From the large numbers of publications regarding artificial insemination, we focused screening on combination keywords namely artificial insemination technology (n=437), artificial insemination materials (n=32), biomaterials AND artificial insemination (n=341), biomaterials AND artificial insemination technology (n=5), and inorganic biomaterials AND artificial insemination technology (n=1). Thus total 816 publications were retrieved using those keywords.

Data related to the study question were extracted and combined with our current research. Data were analysed and synthesized narratively.

## **3. RESULTS AND DISCUSSION**

After assessing the quality of publications through

matching PICOS criteria and risk of bias, we finally selected 65 full papers to further study, as depicted in Table 1.

The 65 selected full papers were matched to criteria through spreading them to the steps of processes and biomaterials used for supporting those steps of artificial insemination processes of ruminant animals. For instance, artificial insemination process of cows [5] that can be described as a flow diagram as detailed on Table 2. The AI processes include animal fertility assessment, semen collection, semen evaluation, semen processing, spermatozoa preservation, and insemination [5]. Those AI steps need supporting biomaterials as well as apparatus/techniques.

Cryopreservation of semen/spermatozoa as the important part of artificial insemination has advantages dan disadvantages as shown in Figure 1.

The advancement of artificial insemination technology in terms of methods/techniques/apparatus and supporting biomaterials reviewed systematically by using literatures since last 5 years. There is a need to develop novel biomaterials as novel procedures to enhance the quality of preserved spermatozoa and

Search keywords	Publication Numbers	Criteria met
"Artificial insemination technology"	437	98
"Artificial insemination materials"	32	16
"Biomaterials" AND "artificial insemination"	341	88
"Biomaterials" AND "artificial insemination technology"	5	3
"Inorganic biomaterials" AND "artificial insemination"	1	1
Total relevant publications after removing duplicates		191
Selected full papers related to artificial insemination materials and process in animal		65
husbandry		

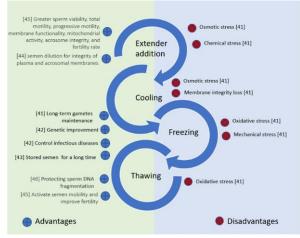


Figure 1. Advantages and Disadvantages of the Cryopreservation Steps.



## Table 2. Steps of AI Processes and Supporting Biomaterials Needed

Steps of AI	Apparatus/Technique/Methods	Supporting Biomaterials	
Processes			
Animal fertility	A sensor based automatic ovulation	enzyme linked immunoassay (ELISA)[6];	
assessment	prediction system [6]; excessive negative	high-density single-nucleotide polymorphism	
	energy balance (eNEB) [7], Genomic data	chip assay [8]	
	[8]; Cow fied data set [9].		
Semen collection	Electroejaculation and transrectal massage	NaCl 0,6% [12]	
	method [10] {16] [17];		
	Artificial vagina equipped with a scaled glass		
	tube [11],		
	collector tube inside the artificial vagina [12]		
Semen evaluation	0,25 mL straws and were sealed thermally	PVA powder [13]; TFYG diluent [20]	
	[13]. CASA Analysis [18][19]; flow cytometric		
	analysis [19];		
	enzyme GOT-GPT leakage pre- and post-		
	freezing [20]		
Semen	Fresh diluted, Equilibration, 0h Post-thaw, 1h	tris egg yolk extender [14] [18],	
processing	Post-thaw [14];	nano-copper and nano-zinc particles during	
	elimination of semen viscosity [22];	in vitro maturation [15];	
	enzymatic and mechanical liquefaction of	DMSA-coated maghemite nanoparticles [21],	
	seminal plasma on freezability of semen [23]	nanoparticle [24]; vitamin [25]	
Spermatozoa	distinct freezing curves after thawing [26];	CeO <sub>2</sub> nanoparticles [27]; plastic straws	
preservation	exposure to CeO <sub>2</sub> nanoparticles during	[28][29][30][31][32]	
	storage at 4 C for 96 hours [27];	[33]; antioxidants reduce lipid peroxidation	
	Cryopreserve in liquid nitrogen [18]	and improve quality of crossbred ram sperm	
	[28][29] [ 30] (31][32][33] [34]	[34]; carboxylated Poly-I-lysine and glycerol	
		on freezability [35];	
		Tris Extender Supplemented with Ethylene	
		Diamine Tetraacetic Acid [36]	
Insemination	Timed Artificial Insemination Following	NaCl to maintain of viability and motility of	
	Oestrous Synchronization [37]; Gedis®	spermatozoa [38]	
	Systems [39];		
	Three-Dimensional (3D) Porcine Preantral		
	Follicles Culture Utilizing Hydrophobic		
	Microbioreactors [40]		

reduce costs as well as an appropriately fit technology for husbandry. For instance, it was reported that the AI for animal husbandry still has problems, especially its effectiveness [41]. Our current works, as purposed in the brief reviews [47] [48], are developing biomaterial supporting artificial insemination based on local raw materials. For instance, the silica biomaterials were originated from rush husk and phosphates of hydroxyapatite from animal bones wastes. Those biomaterials were used to produce a straw having properties of humidity and temperature sensor in the form of silica gel beds in order to develop a novel ambient temperature spermatozoa preservation as well as novel formula of extender for enhancing the quality of semen.



### **4. CONCLUSION**

The current development of AI for animal husbandry has an important role in enhancing productivity, improving the sustainability of food security as well as improving genetic conservation. However, it has still a lack in effectiveness and cost because of cryopreservation needs freezing step using liquid nitrogen. We proposed that developing better techniques and biomaterials supporting AI could be not only low cost but also eco-friendly, which might contribute to the sustainable development of food for all people leading to the human beings as well as animal welfare.

For small tables, please place it within a column and bigger table be placed in a text frame spanning to both columns. Use the Table facility available within the MSWord. The font in the row header should be bold and you can use the style available from the style palette.

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#### REFERENCES

- F. W. Nicholas Genetic improvement through reproductive technology. Animal Reproduction Science. Vol. 42, Issues 1–4, 1996, pp. 205-214, DOI: <u>https://doi.org/10.1016/0378-4320(96)01511-4</u>.
- T. C. Davis & R.R White. Breeding animals to feed people: The many roles of animal reproduction in ensuring global food security. Theriogenology. 2020 Vol. Jul 1, No. 150, pp. 27-33. DOI: <u>https://doi.org/10.1016/j.theriogenology.202</u>0.01.041.
- [3] Centre for Reviews and Dissemination. Systematic reviews: CRD"s 16 guidance for undertaking reviews in health care, York: University of York. 2009. DOI: <u>https://doi.org/0.7748/ns2009.10.24.6.30.p4</u> 606.
- M. Delgado-Rodríguez & M. Sillero-Arenas. Systematic review and meta-analysis, Medicina Intensiva, 2018. Vol. 42, No.7, pp. 444–453. DOI: <u>https://doi.org/10.1016/j.medin.2017.10.003</u>.

- [5] Mulu M, Moges N, Adane M. Review on process, advantages and disadvantage of artificial insemination in cattle. Int J Vet Sci Anim Husb. 2018;3:8-13. <u>https://www.veterinarypaper.com/pdf/2018/</u> vol3issue6/PartA/3-5-5-712.pdf
- T. Mottram, J. Hart & R. Pemberton A Sensor Based Automatic Ovulation Prediction System For Dairy Cows. Sensors and Microsystems, 2000. pp. 44-53. DOI: <u>https://doi.org/10.1142/9789812792013\_000</u> 7
- [7] Macrae A. Assessment of energy balance in dairy cattle. Livestock. 2019 Sep 2;24(5):229-35. DOI: https://doi.org/10.12968/live.2019.24.5.229.
- [8] BS. Olasege, TMS. Mahir, GC. Gouveia, J. IR. Kour, LR. Porto-Neto, BJ. Hayes, BJ, Fortes MR. Genetic parameter estimates for male and female fertility traits using genomic data to improve fertility in Australian beef cattle. Animal Production Science. 2021 Aug 3.https://doi.org/10.1071/AN21097.
- [9] HA. Pacheco, M. Battagin, A. Rossoni, A. Cecchinato, F. Peñagaricano. Evaluation of bull fertility in Italian Brown Swiss dairy cattle using cow field data. Journal of Dairy Science. 2021 Oct 1;104(10):10896-904. DOI: <u>https://doi.org/10.3168/jds.2021-20332</u>
- [10] L.K.Alves, ID. Ávila Caixeta, TF.de Almeida, MM. da Costa, J. França, N.Silva. Influence of the semen collection method on the behavior of cattle. In55a Reunião Anual da Sociedade Zootecnia,  $28^{\circ}$ Brasileira de Congresso Brasileiro de Zootecnia, Goiânia, Brasil, 27 a 30 de agosto de 2018 2018. Sociedade Brasileira de Zootecnia-SBZ, Associação Brasileira dos Zootecnistas. https://www.cabdirect.org/cabdirect/abstract/202

<u>nups://www.cabdirect.org/cabdirect/abstract/202</u> 03172833.

- [11] T. Wahyu Suprayogi, S. Susilowati. The effect of cattle seminal plasma crude protein on the cryopreservation of goat semen. Iranian Journal of Applied Animal Science. 2018 Dec 1;8(4):641-6.
- [12] K. W. Prihatin, L. Hakim, and S. Maylinda Repeatability estimation of semen production and quality of locals Madura cattle breed (Bos Indicus). *Jurnal Kedokteran Hewan*, 1017. *11*(2), pp.70-72. DOI: https://doi.org/10.21157/ j.ked.hewan.v11i2.5942

- E.L. Chung, N. Nayan NS. Nasir, PS. Hing, S. Ramli, MH. Rahman, MH. Kamalludin. Effect of honey as an additive for cryopreservation on bull semen quality from different cattle breeds under tropical condition. J. Anim. Health Prod. 2019;7(4):171-8. DOI:http://dx.doi.org/10.17582/journal.jahp/201 9/7.4.171.178
- [14] Y. Dhaka, M. Singh, A. Sharma, P. Kumar. Establishing a relationship between semen evaluation parameters of freezable and nonfreezable Murrah buffalo bull semen. Indian Journal of Animal Sciences. 2020 Oct 1;90(10): 352–1355.
- [15] B. R. Abdel-Halim, W.A. Moselhy, & N. A. Helmy. Developmental competence of bovine oocytes with increasing concentrations of nanocopper and nano-zinc particles during in vitro maturation. Asian Pacific Journal of Reproduction, 2018. 7(4),
- [16] M. C. Abraham, K. de Verdier, R. Båge & J. M. Morrell Semen collection methods in alpacas. *Veterinary Record*, 2017. 180(25), 613-614.
- [17] S. Al-Bulushi, B.M. Manjunatha, R. Bathgate, J.P. Rickard & S.P. de Graaf. Effect of semen collection frequency on the semen characteristics of dromedary camels. *Animal reproduction science*, 2018. *197*, 145-153.
- [18] Susilawati TR, Ratnawati DI, Isnaini NU, Kuswati K, Yekti AP. Character of liquid semen motility in various diluents on Balinese cattle during cold storage. Asian Journal of Microbiology, Biotechnology and Environmental Sciences. 2018;20(1):166-72.
- [19] Tanga BM, Qamar AY, Raza S, Bang S, Fang X, Yoon K, Cho J. Semen evaluation: methodological advancements in sperm qualityspecific fertility assessment. Asian-Australasian Journal of Animal Sciences. 2021 Apr 23.
- [20] P.J.Chaudhary, A.J. Dhami, D.V. Chaudhari, S.C. Parmar. Freezability of cattle and buffalo semen and association of fresh and frozenthawed sperm quality parameters. Int. J. Curr. Microbiol. App. Sci. 2017. Vol. 6 No. 12. pPp.1445-54.
- [21] D.F. Caldeira, F. Paulini, R.C. Silva, R.B.D. Azevedo & C.M. Lucci. In vitro exposure of bull sperm cells to DMSA-coated maghemite nanoparticles does not affect cell functionality or structure. *International Journal of Hyperthermia*, 2018. Vol. 34 No. 4, pp. 415-422.

- [22] El-Bahrawy, K., Rateb, S., Khalifa, M., Monaco, D., & Lacalandra, G. (2017). Physical and kinematic properties of cryopreserved camel sperm after elimination of semen viscosity by different techniques. Animal reproduction science, Vol. 187, pp. 100-108.
- [23] K. A. El-Bahrawy. Influence of enzymatic and mechanical liquefaction of seminal plasma on freezability of dromedary camel semen. *World's Veterinary Journal*, 2017. Vol. 7 No. 3, pp. 108-116.
- [24] J.M. Feugang, C.E. Rhoads P.A. Mustapha, S. Tardif, J.J. Parrish, S.T. Willard & P. L. Ryan, Treatment of boar sperm with nanoparticles for improved fertility. *Theriogenology*, 2019. Vol. 137, pp. 75-81.
- [25] F. Fitriani, S. Erlina, A. Gunawan, M. Subhan & W. Nugroho. Vitamin e sebagai antioksidan terhadap fertilitas perkawinan ayam Bangkok dengan ayam broiler. *Jurnal Ilmiah Fillia Cendekia*, 2020. Vol. 5. No. 2, pp. 61-64.
- [26] C. E. R. Ferreira, K. L. Goularte, G. D.C. Tavares, S. M. M. Gheller, R.G. Mondadori, A.D. Vieira & T. Lucia. Impact of distinct freezing curves on the quality of ram sperm after thawing. CryoLetters, 2019. Vol. 40 No. 3, pp. 193-199.
- [27] L. Falchi, G. Galleri, G. M. Dore, M. T. Zedda,
  S. Pau, L. Bogliolo, ... & S. Ledda, Effect of exposure to CeO<sub>2</sub> nanoparticles on ram spermatozoa during storage at 4 C for 96 hours. Reproductive Biology and Endocrinology, 2018. Vol. 16 no. 1. Pp. 1-10.
- P. Comizzoli, Separating Good Plastic from Bad Plastic in Conservation Biology. A Chronology of Middle Missouri Plains Village Sites, . 2017. 77.
- [29] P. Comizzoli. Biobanking and fertility preservation for rare and endangered species. *Animal Reproduction (AR)*, 2019. Vol. 14, No. 1. pp.30-33.
- [30] P. Comizzoli. Integrating fertility preservation and cryo-banking into the conservation of rare and endangered deer species. *Animal Production Science*, 2019. Vol. 60. No. 16. pp. 1227-1232.
- [31] P. Comizzoli. Birth of a Giant Panda Cub After Artificial Insemination with Frozen–Thawed Semen: A Powerful Reminder About the Key Role of Biopreservation and Biobanking for Wildlife Conservation. 2020.



- [32] P. Comizzoli & D.E. Wildt. Cryobanking biomaterials from wild animal species to conserve genes and biodiversity: relevance to human biobanking and biomedical research. In *Biobanking of Human Biospecimens*. 2017. pp. 217-235. Springer, Cham.
- [33] Bashawat, M., Hensel, B., Müller, K., & Schulze, M. (2021). Cooled storage of semen from livestock (Part II): camelids, goats, and sheep. Animal Reproduction Science, 106855.
- [34] Banday, M. N., Lone, F. A., Rasool, F., Rashid, M., & Shikari, A. (2017). Use of antioxidants reduce lipid peroxidation and improve quality of crossbred ram sperm during its cryopreservation. Cryobiology, 74, 25-30.
- [35] Akhter, S., Awan, M. A., Arshad, J., Rakha, B. A., Ansari, M. S., & Iqbal, S. (2020). Effect of synergism between carboxylated Poly-1-lysine and glycerol on freezability of Nili-Ravi Buffalo (Bubalus bubalis) Semen. *Biopreservation and Biobanking*, 18(5), 367-375.
- [36] Anwar, R. I., Saili, T., Rizal, M., & Arifiantini, R. I. (2020, March). Quality of Garut Ram's Sperm in Tris Extender Supplemented with Ethylene Diamine Tetraacetic Acid Preserved at Low Temperature (3–5° C). In *IOP Conference Series: Earth and Environmental Science* (Vol. 465, No. 1, p. 012051). IOP Publishing.
- [37] Abdelwahid, H. H., Abdallah, A. A., Mohammed, R. M., Shulukh, E. S. A., & Habib, A. B. (2019). Effects of Timed Artificial Insemination Following Oestrous Synchronization on Pregnancy Rate of Dairy Cattle in the tropics. Int. J. of Multidisciplinary and Current research, 7.
- [38] Agustina, E., Hermadi, H. A., Puntodewo, H., Hernawati, T., Triana, I. N., & Safitri, E. (2017). The Effectiveness of Honey in Physiological NaCl to Maintain of Viability and Motility of Spermatozoa. In Prosiding: The Veterinary Medicine International Conference (Vol. 2017, pp. 609-618). Knowledge E.
- [39] Dimitrov, S., Karapetkovska-Hristova, V., Kochoski, L., Prodanovska-Poposka, V., & Ntsomboh-Ntsefong, G. (2018). Application of Gedis® Systems in Artificial Insemination Technology of Sows. IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT), 12(3), 75-80.
- [40] Duda, M., Gizler, L., & Gorczyca, G. (2021). A Simplified Method for Three-Dimensional (3D) Porcine Preantral Follicles Culture Utilizing

Hydrophobic Microbioreactors. In Next Generation Culture Platforms for Reliable In Vitro Models (pp. 75-84). Humana, New York, NY.

- [41] Ali, S., Iqbal, A., Nawaz, A. H., Yaqoob, A., Zhen, G., ZiNang, J., ... & Zhao, Z. Effects of cryopreservation on Sperm Structural as well as functional Integrity. Sylwan. 2019. Vol. 163, No. 2. pp. 235-276. https://www.researchgate.net/publication/348330 713
- [42] Casali R, Pinczak A, Cuadro F, Guillen-Muñoz JM, Mezzalira A, Menchaca A. Semen deposition by cervical, transcervical and intrauterine route for fixed-time artificial insemination (FTAI) in the ewe. Theriogenology. 2017 Nov 1;103:30-5.
- [43] Parrilla I, Perez-Patiño C, Li J, Barranco I, Padilla L, Rodriguez-Martinez H, Martinez EA, Roca J. Boar semen proteomics and sperm preservation. Theriogenology. 2019 Oct 1;137:23-9.
- [44] Celeghini EC, de Arruda RP, de Andrade AF, Nascimento J, Raphael CF, Rodrigues PH. Effects that bovine sperm cryopreservation using two different extenders has on sperm membranes and chromatin. Animal reproduction science. 2008 Mar 3;104(2-4):119-31.
- [45] Masoudi R, Sharafi M, Shahneh AZ, Kohram H, Nejati-Amiri E, Karimi H, Khodaei-Motlagh M, Shahverdi A. Supplementation of extender with coenzyme Q10 improves the function and fertility potential of rooster spermatozoa after cryopreservation. Animal reproduction science. 2018 Nov 1;198:193-201.
- [46] F. Aghaz, M. Khazaei, A. Vaisi-Raygani, M. Bakhtiyari. Cryoprotective effect of sericin supplementation in freezing and thawing media on the outcome of cryopreservation in human sperm. The Aging Male. 2018 Nov 19.
- [47] E. D. Kusumawati. A brief review on advanced renewable materials for supporting artificial insemination technology. In IOP Conference Series: Materials Science and Engineering. 2021. Vol. 1098, No. 6, pp. 062036. IOP Publishing.
- [48] I. W. Karyasa. Developing renewable thermohydrothermic bioinorganic materials from bone wastes of slaughterhouses. In *Journal of Physics: Conference Series*. 2021, April Vol. 1869, No. 1, p. 012030. IOP Publishing