



# Comparative evaluation of different storage methods of sweet potato (*Ipomea batatas* {L}) and their management

<sup>1</sup>Juliet Nwaneri\*, <sup>1</sup>Chinyere Opara and <sup>2</sup>Thankgod Nwaneri

<sup>1</sup>National Root Crops Research Institute Umudike, Abia State, Nigeria

\*[nwanerijulieta@gmail.com](mailto:nwanerijulieta@gmail.com)

<sup>2</sup>Federal College of Agriculture, Ishiagu, Ebonyi State, Nigeria

## ABSTRACT

*In vivo* and *in vitro* study were conducted to evaluate the efficacy of five storage methods (*wood shaving, use of basket, wood ash, bamboo poles and storage on the floor*) of sweet potato and their management. Diseased tubers of sweet potato were collected from the field trial. The fungus was isolated from diseased potato tuber samples and identified using culture medium. The fungal species associated with the tuber rot of sweet potato and subsequent rot was isolated and identified in this study as *Aspergillus niger*, *Aspergillus flavus*, *Penicilium expansum*, *Rhizopus solanacearum*, *Rhizopus stolonifer*, *Fusarium oxysporium* and *Fusarium solani*. The experiment was carried out between December 2018 and March 2019 for a period of 3 months and repeated in December 2019 to March 2020 to determine the best storage method of sweet potato and their management. From the physical observation the symptom shows three types of rots, which were ascertained as soft, dry and brown rot on Sweet potato tubers in different storage systems. From December 2018 after 3 months of storage were significantly different ( $P < 0.05$ ) on the level of inhibition of the causal organisms on different storage methods used. Wood ash reduced the extent (2.7, 1.8 for dry and soft rot respectively) of tuber rot pathogens more than other storage methods, the least was from the tubers stored on the floor which recorded 95.5 from the dry rot and gave less value in soft rot (6.3), but the tubers stored in basket method had the highest rot incidence (34.8) in soft rot than every other storage method. In 2019 after 3 months storage shows the same sequence followed in 2018. Wood ash still takes the lead in control of rot induced pathogen than other methods with a value of 15.3 from dry rot and 99.53 for the floor which was the least in rot reduction. The degree of rot incidence in 2019 was higher than in 2018. The mean percentage dry rot in 2019 recorded 55.3 while mean dry rot in 2018 was 39.9. For the soft rot, the mean value was 12.4 and 8.7 (2019, 2018) respectively. This could be due to higher temperature regime of drought in 2019. Also, there was appreciable difference in nutrient values of the healthy and infected tubers of sweet potato. The test pathogens significantly affected the nutrient content of sweet potato tubers. *Aspergillus flavus* was the most virulent pathogen depleting the gross nutrient content of the tubers by 48.53%, this was closely followed by *Rhizopus stolonifer* which incurred mean percentage loss of 46.50. However, *Penicilium expansum* was the least virulent in the study. A mean percentage loss of nutrient of 29.65% was recorded on infected tubers due to the pathogen (*P. expansum*).

**Key words:** Diseases, inhibition, microorganism, test plant and treatments

## INTRODUCTION

Sweet potato (*Ipomea batatas*) constitutes an important staple food in the tropics; being one of the major source of carbohydrates, vitamins and dietary fibres. Despite the nutritional and economic importance of sweet potato, its production is limited by pests and diseases. It has been estimated that an average of over 25% of the yield is lost annually to diseases and pests globally (FAO, 1998). One of the major diseases affecting sweet potato production in the tropics is internal microbial rot (PROTA, 2004). Internal rot is the condition that develops in sweet potato tuber where the whole tuber appears wholesome and attractive, but has developed rapid and extensive breakdown of internal tissues (PROTA, 2013). This condition, with time renders the whole tuber rotten. Several microorganisms have been associated with rot in sweet potato. They include; *Aspergillus niger*, *Aspergillus flavus*, *Penicillium expansum*, *Rhizoctonia solanacearum*, *Rhizopus stolonifer*, *Fusarium oxysporium* and *Fusarium solani* (Enyiukwu *et al.*, 2014; PROTA, 2013). Good understanding of the various pests and diseases of sweet potato and their storage management is an important component of improved management technology and produce preservation from microbes-induced spoilages (Salami and Popoola, 2007; Kana *et al.*, 2012). According to Arya (2010), of all losses caused by plant diseases, those that occur after harvest are the most costly. Cassava, yam and sweet potato are important sources of food in the tropics. Others are cocoyam, rice, maize, wheat, sorghum, millet and various fruits, legumes and vegetables. Sweet Potato (*Convolvulaceae; Ipomea batatas*) is an important food crop in Nigeria ranking third amongst important tuber crops of sub-saharan Africa (SSA) after yam and cassava (Amienyo and Ataga, 2007); with 100 million MT produced annually (Ewell and Matuura, 1991; Nwokocha, 1992). The crop according to Oladoye *et al.* (2013) is an important source of anthocyanidines and antioxidants. Rots pose serious problems to its production especially in vegetable potato. In Iran, survey showed that up to 10% pre-harvest and 20% post-harvest rots occurred in the crop (Bidarigh *et al.*, 2012). These rots constitute major impediments to the drives for food security in Nigeria. Therefore, this investigation is aimed at evaluating the efficacy of different storage methods/media on reduction of fungi-induced rots of sweet potato; and on the improvement of the storageability of the tubers in the humid tropical agro-ecology of Umudike, Nigeria. The specific objectives are to:

1. Isolate and identify the fungal rot organisms associated with stored sweet potato in Umudike.
2. Identify the most suitable low-input storage methods/ media for improved shelf-life of the tubers in the agro-ecology
3. To investigate the mechanism of damage by the pathogenic organisms during pathogenesis (Histological studies).

## **MATERIALS AND METHODS**

This study was conducted at the NRCRI, Umudike, Nigeria. Relatively disease – free sweet potato were collected from the Sweet potato programme field trials of the National Root Crops Research Institute, Umudike. The treatments were as follows: wood shavings and wood ash, *use of basket, storage on the floor* and bamboo poles were obtained from the Institute neighborhood. The experiments were set up both *in vivo* and *in vitro*.

### **In Vivo experiment**

The shelf – life of sweet potato tubers were evaluated by the *use of basket, storage on the floor, raised bamboo plat forms, wood shaving and wood ash*. Under shade storage consist of tubers stored on wood ash, wood shavings, raised bamboo poles and basket. Twenty (20kg) kilograms of the relatively diseased free tubers were stored on each storage method. The experiment was carried out in 2018 and 2019 for a period of 3 months. Data taken was on the types and number of rotted and un-rotted tubers per storage method/ medium per replicate.

### **In Vitro experiment**

Rotted tubers were enveloped and taken to the Plant Protection Laboratory NRCRI, Umudike. Infected tubers were washed into water and rinsed in several changes of 500mL sterile distilled water. Bits of the diseased tuber tissues were plated on solidified synthetic PDA (Melbonne, Germany) contained in Petri dishes. The dishes were incubated for 7 days at room temperature of 27°C. The mycelia that grow out of the tuber were repeatedly sub-cultured on fresh 20mL solidified PDA to obtain pure cultures of the organism. Bits of the 10-day old pure stocks were lifted unto a drop of lactophenol in cotton blue on sterile slides, teased gently and cornered with a cover slip. The slides thus prepared were mounted on a microscope and the identity of the organism ascertained with reference to Barnett and Hunter (1995).

### Statistical analysis

Data generated from the experiment were analysed by ANOVA using GENSTAT 2008 Version. Means was separated by Fishers LSD (FLSD) at 0.05 Probability.

### Results and Discussion

This study shows different types of rot infection on sweet potato tubers in different storage system. From the physical observation the symptom shows three types of rots, which were ascertained as soft, dry and brown rot. The result from Table 1 shows that a total of seven micro-organisms (*Aspergillus niger*, *Aspergillus flavus*, *Penicilium expansum*, *Rhalstonia solanacearum*, *Rhizopus stolonifer*, *Fusarium oxysporium* and *Fusarium solani*) were isolated from the rotted tubers of sweet potato, their frequency of occurrence and types of rots they cause are shown below.

**Table 1: Microorganisms isolated from Sweet potato, their frequency of occurrence and type of rots**

Fungi isolate	Frequency of occurrence (%)	Types of rots
<i>Aspergillus niger</i>	13.7	Dry rot
<i>Aspergillus flavus</i>	10.9	Dry rot
<i>Rhizopus stolonifera</i>	60.1	Soft rot
<i>Rhalstonia solanacearum</i>	57.3	Brown rot
<i>Fusarium oxysporium</i>	12.6	Dry rot
<i>Peniciliumexpansium</i>	28.0	Dry rot
<i>Fusarium solani</i>	8.2	Dry rot

Table 2 shows the effect of storage methods on mean rots of *Ipomea batatas* in 2018 after 3 months of storage. There were significant differences ( $P < 0.05$ ) on the level of inhibition of the causal organisms on different storage methods used. Wood ash reduced the extent of (2.7, 1.8 for dry and soft rot respectively) tuber rot induced pathogens more than other storage methods used and the least was from the tubers stored on the floor which recorded 95.5 from the dry rot and gave less value in soft rot (6.3), but the tubers stored in basket method had the highest rot incidence (47.2) in soft rot than every other storage method.

**Table 2: Effect of storage methods on mean rots of *Ipomea batatas* in 2018 (3 Months of storage)**

Storage methods	Dry rot (%)	Soft rot (%)	Brown rot (%)
Bamboo	41.8	5.5	4.0
Floor	95.5	6.3	21.8
Basket method	47.2	20.8	26.2
Wood ash	2.7	1.8	1.0
Wood shaving	12.2	9.0	5.8

Means	39.9	8.7	11.8
<b>LSD0.05</b>	<b>16.1*</b>	<b>4.7*</b>	<b>NS</b>

Table 3 also shows the effect of storage methods on mean rots of *Ipomea batatas* in 2019 after 3 months storage. The same sequence followed in 2018. Wood ash still takes the lead in control of rot induced pathogen in tuber rot of sweet potato than other methods with a value of 7.5 from dry rot and 99.3 for the floor which was the least in rot reduction in storage. But soft and wet rot were not significant. There were significant differences ( $P < 0.05$ ) in the two years experiment. The degree of rot incidence in 2019 was higher than in 2018. The mean percentage dry rot in 2019 recorded 55.3 while mean dry rot in 2018 was 39.9. For the soft rot, the mean value were 12.4 and 8.7 (2019, 2018) respectively. This could be due to higher temperature regime of drought in 2019.

**Table 3: Effect of storage methods on mean rots of *Ipomea batatas* in 2019 (3 Months of storage)**

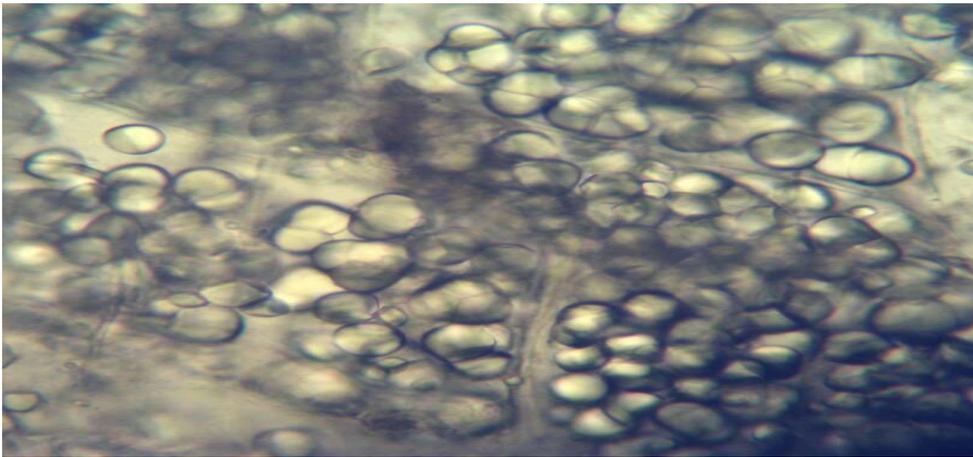
Storage methods	Dry rot (%)	Soft rot (%)	Brown rot (%)
Bamboo	87.3	9.3	8.0
Floor	99.3	7.5	18.8
Basket method	47.2	26.5	28.5
Wood ash	8.7	3.83	3.8
Wood shaving	33.8	14.7	12.5
<b>Means</b>	<b>55.3</b>	<b>12.4</b>	<b>14.3</b>
<b>LSD0.05</b>	<b>13.2**</b>	<b>3.1**</b>	<b>NS</b>

The results in Table 4 indicated appreciable difference in nutrient values of the healthy and infected tubers of sweet potato. The pathogens significantly affected the nutrient content of sweet potato tubers. *Aspergillus niger* was the most virulent pathogen depleting the gross nutrient content of the tubers by 48.53%, this was closely followed by *Rhizopus stolonifer* which incurred mean percentage loss of 46.50. However, *penicilium expansum* was the least virulent of the pathogens affecting the tubers in the study. A mean percentage loss of nutrient of 29.65% was recorded on infected tubers due to the pathogen (*P. expansum*). These pathogen organisms that reduced the nutrient content of tubers has been reported by Amadioha, 2004 which is in agreement with the result of this study.

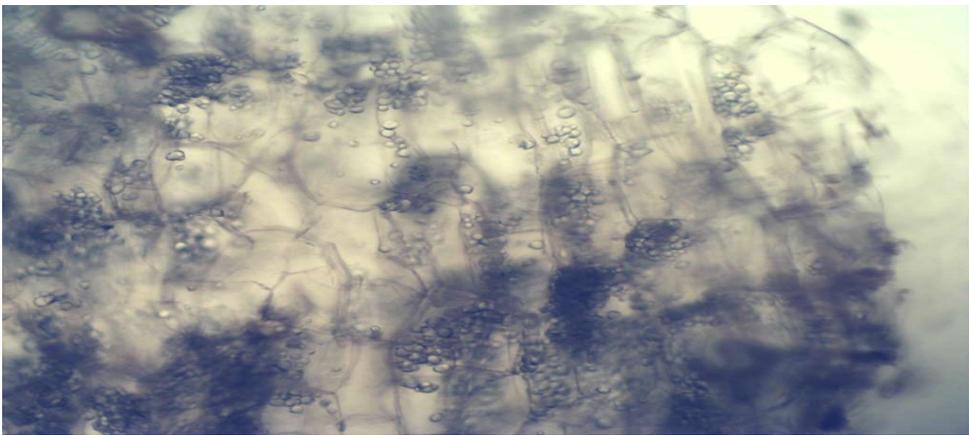
**Table 4: Depletion of nutrient content of Sweet potato tuber by the pathogenic organism at**

Umudike					
Pathogen	Healthy	Infected	Loss	%Loss	Mean% loss
<i>A. niger</i>	11.87	6.11	5.76	48.53	18.11
<i>R. stolonifer</i>	11.87	6.35	5.52	46.50	17.56
<i>F. oxysporum</i>	11.87	7.62	4.25	35.80	14.89
<i>P. expansum</i>	11.87	8.35	3.52	29.65	13.35
Mean	11.87	7.11	4.76	40.10	5.09
<b>LSD (0.05)</b>	<b>NS</b>	<b>1.64</b>	<b>0.38</b>	<b>1.02</b>	<b>1.01</b>

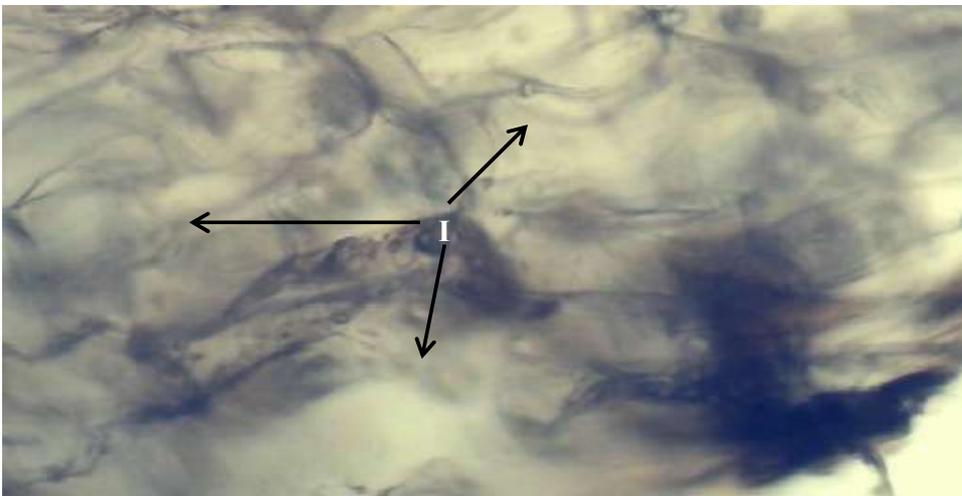
**PLATE A – E SHOWS DEPLETION OF STARCH GRAINS OF SWEET POTATO TUBER BY DIFFERENT FUNGAL ORGANISMS**

X  
1  
0  
0

a

X  
1  
0  
0

b

X  
1  
0  
0

c

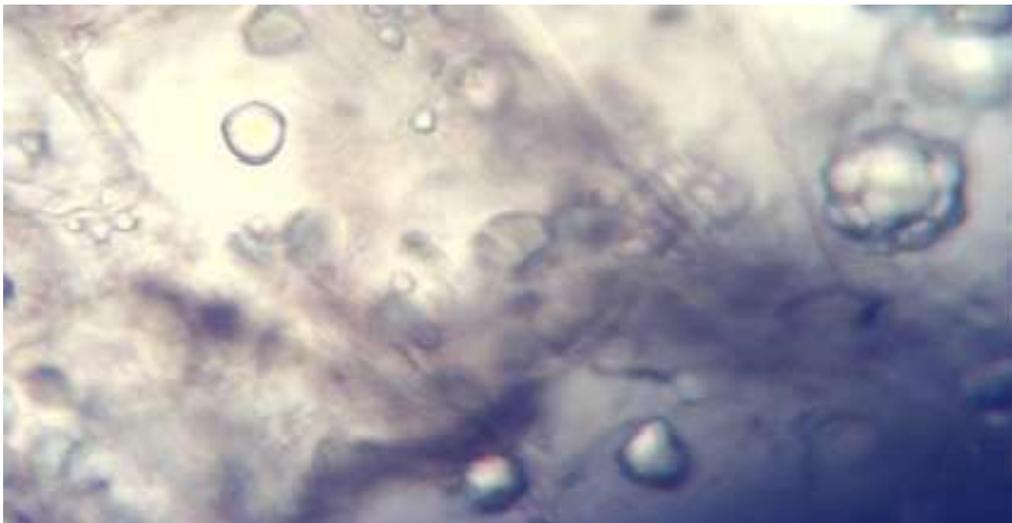
- (a) Fresh healthy sweet potato (*Ipomea batatas*) showing intact cells packed with starch grains.  
 (b) Cells depleted of starch grains 1 week after infection by *Aspergillus niger*  
 (c) Total collapse of cell wall boundaries 2 weeks after infection by *Aspergillus niger*. i: shows depletion of starch grains and ramification of hyphae both inter and intracellular tissue completely macerated.



X  
1  
0  
0

d

(d) Total collapse of cell boundaries and ramification of hyphae both inter- and intracellular 2 weeks after infection by *Rhizopus stolonifer*



X  
1  
0  
0

e

(e) Depletion of cell wall boundaries and ramification of hyphae both inter- and intracellular 2 weeks after infection by *Penicillium expansum*

### Conclusion

This study indicates that there was greater occurrence of dry rot than soft or brown rots amongst the storage methods used. It also shows that wood ash compared well in rot control than other treatments. This may be as a result of wood ash being alkaline in nature and could be that the organisms do not operate well in an alkalinity environment. Therefore, wood ash is a good storage media for *I. batatas* and can effectively minimize post-harvest rot development of Sweet potato tubers in storage. While storing in cemented floor is not an ideal storage media for Sweet potato because it encourages rot development especially dry rot. The reason could be that cemented floor absorbs heat more than other methods in this study.

### References

Amadioha A. C. (2004). Control of black rot of potato caused by *Rhizoctonia*

- bataticola* using some plant leaf extracts. Arch. Plant Pathol. Plant Protect. 37:111-117.
- Amienyo CA, Ataga AE, 2007. Use of indigenous plant extracts for the protection of mechanically injured sweet potato (*Ipomea batatas* (L.) Lam) tubers. Sci Res Essay, 2(5):167-170
- Arya A, 2010. Recent advances in the management of plant pathogens: Botanicals in the fungal pest management. In: Management of fungal plant pathogens. Arya A. and Perello, A. E. (Eds). UK, CAB International, pp. 1-25.
- Bidarigh S, Massiha A, Mohamme RM, Khoshkholgh P, Issazadeh K, Muradov PZ, Azapour E, 2012. Antimicrobial (screening) properties of various plant extracts from *Ocimum basilicum* L. and *Nerium oleander* L. against common fungal organism of potato. J Basic Appl Res, 2(7):6810-6815.
- Ewell PT, Matuura J, 1991. Tropical root crops in a developing economy. Proceeding of the 9th symposium of the International Society for Tropical Root Crops 20-26 October 1991 Accra Ghana. Pp. 405.
- FAO, (1998). Storage and Processing of roots and tubers in the tropics. Rome. Italy. FAO Corporate Document Repository Agro. Industries and Post-harvest Management Services.
- Kana HA, Aliyu IA, Chammang HB, 2012. Review on neglected and underutilized root and tuber crops as food security in achieving the millennium development goals in Nigeria. J Agric Vet Sci, 4:27-33.
- Nwokocha HN, 1992. Agronomy of sweet potato. Root Crop Research and Technology Transfer Training Course (Manual) NRCRI, pp77-84
- Oladoye CO, Olaoye OA, Cornnerton IF, 2013. Isolation and identification of bacteria associated with spoilage of sweet potatoes during postharvest storage. Int J Agric Food Sci, 3(1):10-15.
- PROTA (2004). Vegetable in: Plant Resources of Tropical Africa. PROTA CTA Wageningen, Netherland p 667
- PROTA (Plant Resources of Tropical Africa) (2013). *Solenostemon rotundifolius* Poir (Synonyms: *Germania rotundifolius* Poir, *Plectranthus roundifolius* Sims) database. PROTA.org/db/w-wpd/exec/db Retrieved November
- Salami OA, Popoola OO, 2007. Thermal control of some postharvest rot pathogens of Irish potato (*Solanum tuberosum* L.). J Agric Sci, 52(1):17-31.

**Open Access** This chapter is licensed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits any noncommercial use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license and indicate if changes were made.

The images or other third party material in this chapter are included in the chapter's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the chapter's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.

