

## The Perturbation Toxic Effect of Graphene on HepG2 Cell

Guozheng Jiao<sup>1a</sup>, Xin Li<sup>2a,b\*</sup>, Junqiang Qiu<sup>3c</sup>, Ning Zhang<sup>4c</sup>, Hongying Xu<sup>5c</sup> and Shumin Liu<sup>6,c\*</sup>

<sup>1</sup> Department of Chemistry, Harbin Institute of Technology, Harbin 150090, China

<sup>2</sup> State Key Lab of Urban Water Resource and Environment, Harbin Institute of Technology, Harbin 150090,

<sup>3</sup> Institute of Traditional Chinese Medicine, Key Laboratory of Chinese Materia Medica, Ministry of Education, Heilongjiang University of Chinese Medicine, Harbin 150040, China

<sup>a</sup> E-mail:lixin@hit.edu.cn, <sup>c</sup> E-mail:keji-liu@163.com,

\* the corresponding author

**Keywords:** graphene, HepG2, metabolomics, toxicity

**Abstract.** How to accurately and comprehensively unveil the toxic effect of graphene on the cells have always received extensive attention by many researchers. Owing to the opposition results in classical assay approaches, some researchers have attempted to adopt advanced analysis techniques. Our group successfully performed mass spectrometry-based metabolomics to investigate the toxic effect of graphene on the HepG2. In this analysis platform, by pattern recognition and metabolism analysis, we acquired twelve potential biomarkers and two main disturbed metabolism pathways involving in glutathione metabolism and arginine and proline metabolism. Our findings demonstrated that the graphene stimulates the HepG2 cell and results in breaking the homeostasis and perturbing the metabolism pathway, we should focus on this disturbance from the graphene.

### Introduction

Graphene, and its derivatives, owing to having the unique chemical and physical property, are widely studied, manufactured, as well as used, not only in photoelectricity physical field, but also in biomedical scope. So the chance of exposure of it for the common people are more and more pronounced, the potential health risk and hazard are focused by people. Although the classical *in vitro* toxic assay methods are popular in testing the purity compounds in biomedical, many researchers still have found that some assay methods *in vitro* are disturbed by various modes in testing nanomaterials, such as, disturbance of the assay agents including adsorption or/and stimulus redox, interference in detection optical route, invoking cells inactive sensitivity assay agents and so on. The graphene nanomaterials have also been found disturbing some assay approaches by reported similar pattern. Therefore, *in vitro* assay too many results were revealed the inconsonant and irrelevant and aroused too much controversy about the toxic or no-toxic of the graphene nanomaterials.[1] Some researchers have attempted to adopt advanced analysis techniques to assay the toxicity of graphene nanomaterials. Previously, we have successfully performed the metabolomics to assess the toxic effect of water soluble graphene on the HepG2 cell.[2] Hence, once again we carried out the metabolomics platform to investigate the toxicity of graphene on HepG2.

In the current study, we adopted the UPLC-Q-TOF-MS to assay the metabolites of HepG2 cells treated with graphene, then combined the pattern recognition with the biological events to detailedly expound the disturbance metabolites and perturbation metabolism pathway. We acquired twelve metabolites and two main pathways disturbed by graphene including glutathione metabolism and arginine and proline metabolism. It demonstrated that the graphene has some extent disturbance for the HepG2 cell, *in vivo* usage, some great attention should be taken.

## Results and Discussion

### Pattern recognition

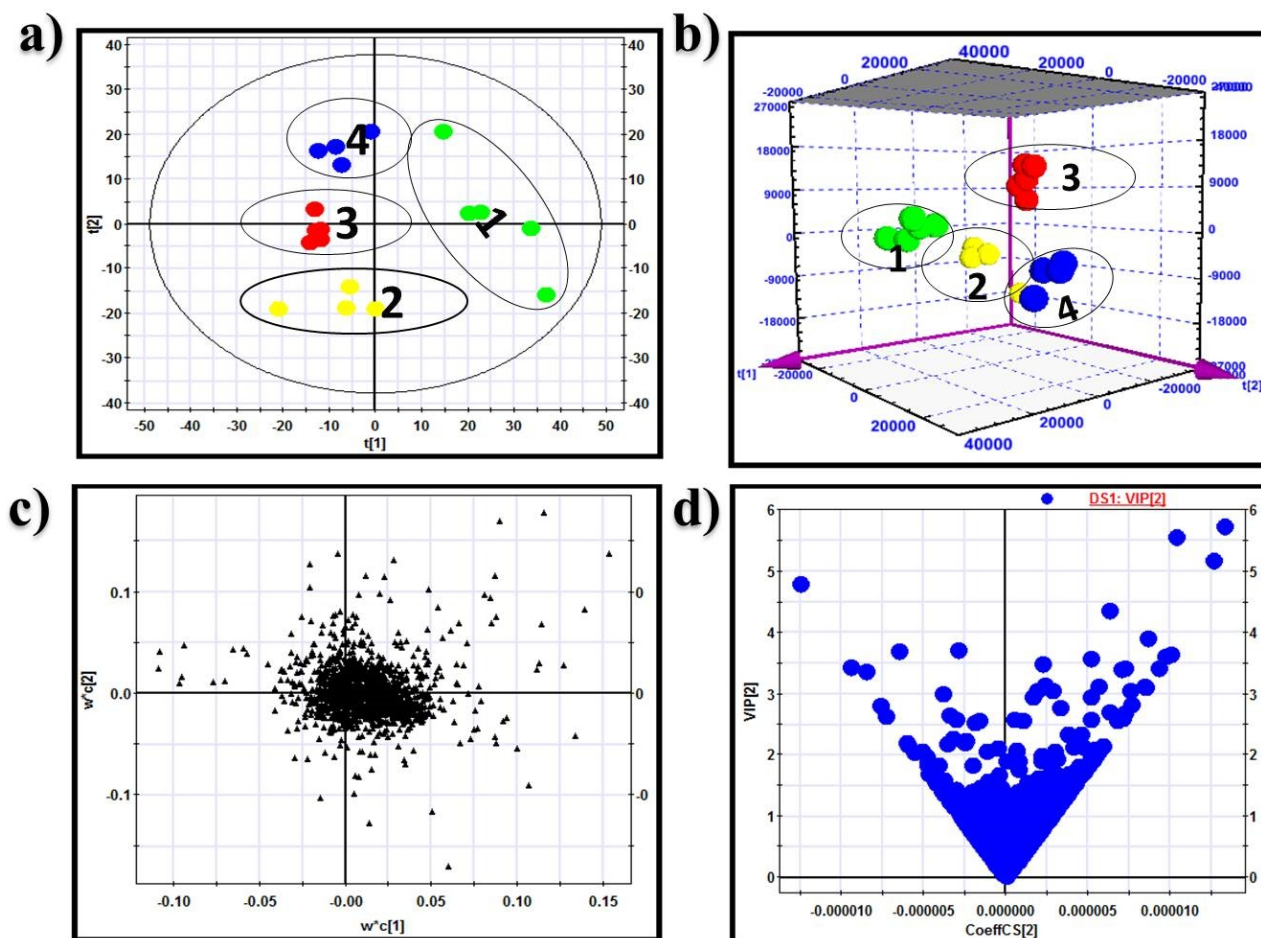


Fig. 1 The metabolic profile of Chang liver cell treated with graphene. *a*). the score plot of PCA; *b*). 3D-score plot of PLSD\_A; *c*). Loading plot of PLSD\_A; *d*). The VIP score plot of PLSD\_A. *circle1*: control dose group (PBS); *circle2*: low dose group (graphene,  $5 \mu\text{g}\cdot\text{mL}^{-1}$ ); *circle3*: middle dose group (graphene,  $40 \mu\text{g}\cdot\text{mL}^{-1}$ ); *circle4*: high dose group (graphene,  $1.00 \mu\text{g}\cdot\text{mL}^{-1}$ ).

All the mass data were input the masslynx 4.1(Waters, USA) and Ezinfo 2.0 software (Waters, USA) and dealt with the multi-analysis including the principal component analysis (PCA) and partial least squares-discriminate analysis (PLS-DA), and we acquired the HepG2 cell metabolic profile (in Fig. 1). In Fig.1a, the PCA score plot, we can find that there is clearly clustering and distinguishing between the treated groups (circle 2, 3, and 4) and the control groups (circle1); in Fig. 1b, the 3D PLSD-A score plot, the separation trending was further magnified. So, it demonstrated that the graphene could disturb the metabolic pathway of HepG2 cells (even low dose  $25 \mu\text{g}\cdot\text{mL}^{-1}$ ), from the low dose to the high dose, the metabolism trajectory of HepG2 cell displayed the clockwise change and the pronounced dose-dependent. Combined the loading plot (Fig. 1c) with VIP score plot (Fig. 1d), we acquired 336 potential biomarker and threshold value setting  $\text{VIP} > 1$ .

The potential biomarkers were suffered from the the MS/MS analysis by the precursor ion scan and retention time, then the mass data in integrated with the free database METLIN, metabolite, ChemSpider, HMDB, MassBank and KEGG to identify the structure of the potential markers (table1). The 12 metabolites are most of the nitrogen-containing compound including amino acid and its derivative, small molecule peptide, and so on

## Univariate analysis

Table 1 the identified markers

Code	Rt	[M+R]	Data	Formula	Name
M1	3.15	[M+H] <sup>+</sup>	307.323	C10H17N3O6S	Glutathione
M2	2.78	[M+H] <sup>+</sup>	252.2267	C10H12N4O4	Deoxyinosine
M3	6.75	[M+H] <sup>+</sup>	488.324	C14H26N4O11P2	Citicoline
M4	0.56	[M+H] <sup>+</sup>	115.1305	C5H9NO2	Proline
M5	0.97	[M+H] <sup>+</sup>	145.2459	C7H19N3	Spermidine
M6	1.60	[M+Li] <sup>+</sup>	184.0431	C6H11NOS2	Sulforaphane
M7	0.53	[M+H] <sup>+</sup>	221.0320	C6H9N2O5P	3-(Imidazol-4-yl)-2-oxopropyl
M8	3.62	[M+H] <sup>+</sup>	184.0634	C5H13NO4S	Unkown
M9	4.19	[M+H] <sup>+</sup>	226.1192	C10H15N3O3	1-(Methylnitrosoamino)-4-(3-pyridinyl)-1,4-but anediol
M10	2.13	[M+Na] <sup>+</sup>	275.0689	C16H12O3	1-[2-(4-Hydroxyphenyl)-1-benzofuran-3-yl]eth anone
M11	8.39	[M+ACN+H]	804.4819	C44H42N8O5	Trp Trp Trp Trp
M12	7.96	[M+H] <sup>+</sup>	466.1759	C20H27N5O6S	Unkown

The signal intensity of potentials biomarkers were dealt by the univariate analysis (analysis of variance). All the change trending of metabolites were displayed by the box-figure (Fig.2), we can find that signal intensity of five metabolites (M2, M6, M7, M10, M11) are decreasing with the graphene concentration, that is the signal intensity of metabolite is inversely proportional to the added graphene dose, in contrast, that of the others (8 metabolites) are increasing with the graphene concentration. It demonstrated that the markers could were intensively disturbed by the graphene. The significant differences values were acquired by virtue of all the treated groups compared with the control group.

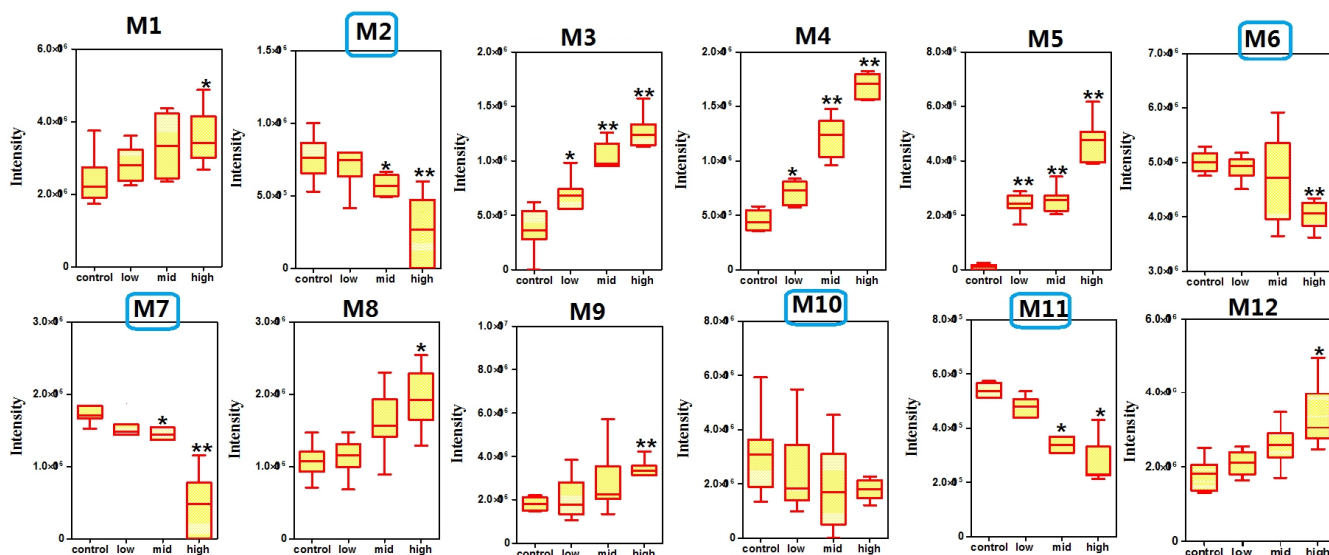


Fig.2 The Box-Figure univariate analysis. Statistical significance were displayed by compared the treated group (Low, Middle, and High dose group) with the Control group. Significant difference stands as “\*”  $p < 0.05$ , and “\*\*”  $p < 0.01$ .

## Pathway analysis

All the biomarkers were input the KEGG, HMDB, MetPA, and SMPD database, we obtained the the disturbed metabolism pathway of HepG2 treated with graphene[3]. According to the pathway impact value (setting threshold value  $> 0.1$ ), we choose the disturbed metabolism pathway glutathione

metabolism and arginine and proline metabolism (Fig. 3). The two disturbed metabolism pathway were disturbed by glutathione (M1), proline(M4), and spermidine(M5). In Fig. 2, we can find that the changed trending of these 3 markers, the glutathione, proline, and spermidine are also displayed increasing with the graphene dose.

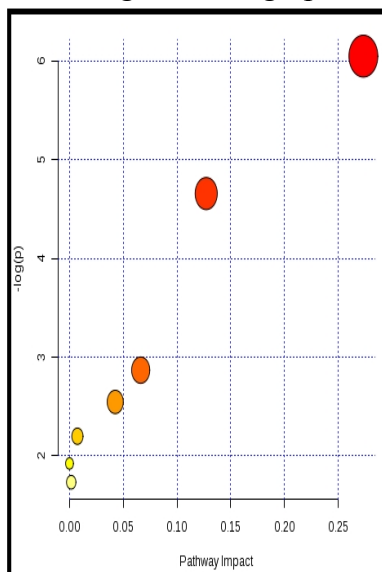


Fig. 3 pathway impact

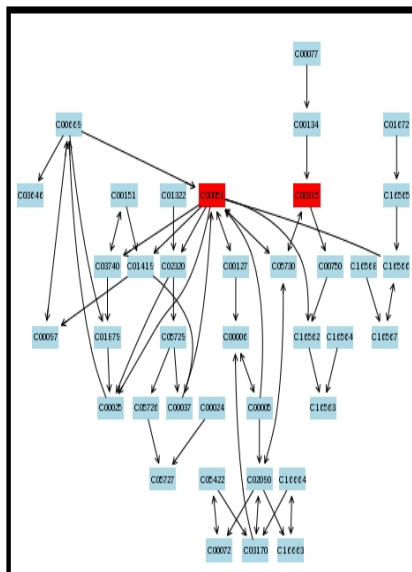


Fig. 4 Glutathione metabolism

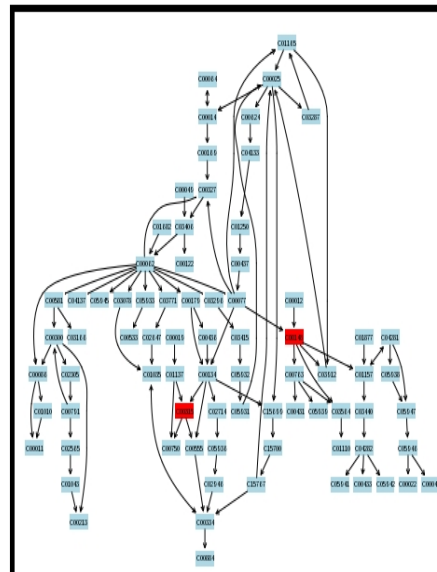


Fig. 5 arginine and proline metabolism

The two pathways of glutathione metabolism (Fig. 4) and arginine and proline metabolism (Fig. 5) play an important role in cell living activities.[4] Glutathione is mainly used to deal with the toxic waste and help detoxify, the crucial thiol (-SH) group play an important role in antioxidant. In liver, it can be used to conjugate reaction with the toxic chemicals and dispose them for detoxify. Additionally, the cell can help construct the red and white cell and improve the immunity. It can also be used to clinical to reduce the damages of the chemotherapy and radiation in cancer treatments, and can reduce the toxicity of heavy metal. Glutathione can also help the conjugation reactions and reduction reactions in in cytosol, microsomes, and mitochondria, and so on. The proline can be used forming the protein, moreover, it plays an important role in constructing the collagen. The pathway of arginine and proline metabolism is involved in the urea cycle. Spermidine is thought to improve the stabilize of some membranes and nucleic acid structures. In Fig. 3, Fig.4, and Fig.5, we can find that the homeostasis of the mentioned pathways was disturbed, so the HepG2 biological balance may be perturb by graphene. We should focus on this perturbation before widely usage this kind of nanomaterials. The further mechanism is still needed to be done.

## Summary

In this study, we can find that the metabolites of HepG2 cells were disturbed by graphene, and two metabolism pathways including the glutathione metabolism and arginine and proline metabolism were disturbed by graphene. So, in order to make good use of the graphene nanomaterials, we should focus on these possibility perturbations of cells.

## References

- [1] G.X, M. N. Assessment of the toxic potential of graphene family nanomaterials. Journal of food and drug analysis,22 (2014) 105-115.
- [2] J. GZ, Li X, Zh. N, Q. J.Q, X. H.Y., L. SM. Metabolomics study on the cytotoxicity of graphene. Rsc Adv, 4(2014) 44712-7.
- [3] X.J, S.I.V, Han B, Wishart DS. MetaboAnalyst 3.0-making metabolomics more meaningful. Nucleic acids research43 (2015) 251-257.

[4] W. DS, F. A, K. C, et al. SMPDB: The Small Molecule Pathway Database. *Nucleic Acids Res.* 38(2010) (Database issue) 480-487.