

Preparation of Diuridine and Dicytidine Pentaphosphates

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Abstract. Two symmetrical dinucleoside pentaphosphates, Up₅U and Cp₅C, were synthesized via the activation of the P (V)–N bond. The key nucleoside phosphoropiperidates intermediate were obtained by a redox condensation method. The reaction of the phosphoropiperidate with uridine and cytidine tetraphosphate in the presence of 4, 5-dicyanoimidazole (DCI) afforded the desired dinucleoside pentaphosphates in moderate isolated yields.

Introduction

Dinucleoside polyphosphates, or dinucleotides (Np_nN' N, N'=A, U, G, C; n = 2–7) are naturally occurring compounds [1]. They consist of a polyphosphate chain linked to two nucleosides at the 5'-positions on both ends. Many important functions have been elucidated for Ap_nAs, Ap_nGs and Gp_nGs in various biological processes, such as platelet disaggregation, neurotransmission, modulation of vascular tone, and cell signaling [2–5]. They have also been shown to act as substrates for a variety of DNA polymerases and function as inhibitors of kinases, endoribonuclease, IMP dehydrogenase, and adenylosuccinate synthetase. In addition, the Ap₄A may be used as primers for DNA synthesis by DNA polymerase or for RNA polymerase, and the Ap₅A–Zn²⁺ complex have been used to reveal preferred pathways that create the configuration capable of proficient chemistry in the catalysis of adenylate kinase [6–10]. Due to the chemical and metabolic stability of dinucleotides, they are attractive as therapeutic agents. Several Np_nN' analogues have been administered in human clinical trials [11–13]. For instance, Ap₄A has been tested for lowering blood pressure during anesthesia. Artificial dinucleoside polyphosphates Up₄U and Up₄dC have been developed for the treatment of dry eye disease and cystic fibrosis, respectively.

Currently, the method for the preparation of dinucleoside pentaphosphates is still very limited and facile synthesis of dinucleoside pentaphosphates remains a challenge for phosphorus chemists. The most commonly used method is the approach based on the synthesis of dinucleoside di-, tri-, and tetraphosphates. The direct coupling of nucleoside diphosphates (NDPs) and nucleoside triphosphates with excess condensing reagents such as carbonyldiimidazole (CDI) or dicyclohexylcarbodiimide (DCC) could only afforded the desired products in low yields [14–16]. Recently, dinucleoside pentaphosphates including Ap₅A, Gp₅G, and Ap₅G have been efficiently synthesized by coupling nucleoside 5'-tetraphosphate with nucleoside 5'-phosphoromorpholidates with 4,5-dicyanoimidazole activator. On the basis of this approach, we report in this paper the synthesis of dicytidine and diuridine pentaphosphates from nucleoside

phosphoropiperidate and corresponding nucleoside tetraphosphates via the activation of P (V)–N bond of the phosphoropiperidate precursors.

Experimental

All reactions were performed in anhydrous solvents under an atmosphere of dry argon. The triethylammonium salts of nucleoside 5'-phosphoropiperidates were synthesized according to the procedure described in a previous report [17]. Nucleoside 5'-tetraphosphates was prepared according to a known method. Ion exchange chromatography employed DEAE Sephadex A-25 exchanger. Preparative HPLC was equipped with a RP C18 column (19 mm 250 mm, 10 μ m). NMR spectra were obtained with a 400 MHz instrument with chemical shifts reported in parts per million (ppm, δ) and referenced to D₂O. IR spectra were recorded on a FT-IR spectrometer. Low-resolution mass spectra were obtained with an ion trap mass spectrometer and reported as *m/z*.

***P*¹, *P*⁵-Dicytidine-5', 5'-Pentaphosphate, Pentasodium Salt (5)**

To a solution of cytidine 5'-phosphoropiperidate (1) (49 mg, 0.1 mmol) in *N*-methylpyrrolidone (2 mL) were added cytidine 5'-tetraphosphate (3) (tetra-*n*-butylammonium salt, 61 mg, 0.04 mmol) and DCI (24 mg, 0.2 mmol). The reaction was stirred at 20 °C for 20 h. The white precipitation was collected by centrifuge. The crude product was dissolved in deionized H₂O (0.5 mL) and loaded on a DEAE Sephadex A-25 ion exchange column (1.6 cm 25 cm). Elution with NH₄HCO₃ buffer (linear gradient 0.5 to 0.9 mol/L), combination of appropriate fractions, and lyophilization afforded dinucleoside pentaphosphate in ammonium salt form. To remove the small amount of contaminated polyphosphate byproducts, the ammonium salt was further purified by a preparative RP HPLC [flow rate = 20 mL/min; linear gradient of 0–10% MeOH in TEAB buffer (10 mmol/L, pH 8.0) over 15 min; UV detection at 254 nm]. Combination of appropriate fractions and lyophilization afforded dinucleoside pentaphosphate in triethylammonium salt form. Passage of the solution of the triethylammonium salt in deionized H₂O through a bed of Dowex 50W-X8 ion exchange resin (Na⁺ form) and lyophilization afforded **5** (14 mg, 35%) as pentasodium salt. ¹H NMR (400 MHz, D₂O): δ 7.84 (d, *J* = 7.2 Hz, 2H), 6.11 (d, *J* = 7.2 Hz, 2H), 5.96–5.91 (m, 2H), 4.34–4.26 (m, 4H), 4.25–4.22 (m, 4H), 4.18–4.15 (m, 2H) ppm; ¹³C NMR (100 MHz, D₂O): δ 166.1, 157.7, 141.2, 96.5, 88.3, 82.6, 75.4, 69.3, 65.8 ppm; ³¹P NMR (162 MHz, D₂O): δ -11.5 (m, 2P), -22.9 (m, 3P); IR (KBr): ν_{max} 3697, 3443, 2920, 2580, 2353, 1658, 1601, 1529, 1495, 1409, 1291, 1242, 1120, 1080, 948, 810, 600, 516 cm⁻¹; LRMS (ESI⁻): *m/z* calcd for C₁₈H₂₉N₆O₂₄P₅ [M–H]⁻ 868.0; found 868.1.

***P*¹, *P*⁵-Diuridine-5', 5'-Pentaphosphate, Pentasodium Salt (6)**

To a solution of uridine 5'-phosphoropiperidate (2) (49 mg, 0.1 mmol) in *N*-methylpyrrolidone (2 mL) were added uridine 5'-tetraphosphate (4) (tetra-*n*-butylammonium salt, 61 mg, 0.04 mmol) and DCI (24 mg, 0.2 mmol). The reaction was stirred at 20 °C for 18 h. The white precipitation was collected by centrifuge. The crude product was dissolved in deionized H₂O (0.5 mL) and loaded on a DEAE Sephadex A-25 ion exchange column (1.6 cm 25 cm). Elution with NH₄HCO₃ buffer (linear gradient 0.5 to 0.9 mol/L), combination of appropriate fractions, and lyophilization afforded dinucleoside pentaphosphate in ammonium salt form. To remove the small amount of contaminated polyphosphate byproducts, the ammonium salt was further purified by a preparative RP HPLC [flow rate = 20 mL/min; linear gradient of 0–10% MeOH in TEAB buffer (10 mmol/L, pH 8.0) over 15 min; UV

detection at 254 nm]. Combination of appropriate fractions and lyophilization afforded dinucleoside pentaphosphate in triethylammonium salt form. Passage of the solution of the triethylammonium salt in deionized H₂O through a bed of Dowex 50W-X8 ion exchange resin (Na⁺ form) and lyophilization afforded **5** (15 mg, 38%) as pentasodium salt. ¹H NMR (400 MHz, D₂O): δ 7.92 (d, *J* = 8.0 Hz, 2H), 5.95–5.92 (m, 4H), 4.38–4.32 (m, 4H), 4.23–4.20 (m, 6H) ppm; ¹³C NMR (100 MHz, D₂O): δ 166.4, 152.1, 141.8, 102.9, 88.2, 83.6 (d, *J* = 8.9 Hz), 73.8, 69.8, 65.1 (d, *J* = 5.2 Hz) ppm; ³¹P NMR (162 MHz, D₂O): δ -11.0 (m, 2P), -22.4 (m, 3P) ppm; IR (KBr): ν_{\max} 3870, 3749, 3690, 3457, 2923, 2741, 2332, 1605, 1481, 1388, 1336, 1306, 1247, 1117, 1079, 954, 825, 721, 649, 512 cm⁻¹; LRMS (ESI⁻): *m/z* calcd for C₁₈H₂₇N₄O₂₆P₅[M-H]⁻ 870.0; found 870.1.

Results and Discussion

As shown in Fig. 1, cytidine phosphoropiperidate (**1**) and uridine phosphoropiperidate (**2**) were treated with 0.4 equiv of cytidine tetraphosphate (**3**) and uridine tetraphosphate (**4**), respectively. In the presence of 2 equiv of DCI at 20 °C in *N*-methylpyrrolidone, the reactions were stirred for 18–20 h. We found that dinucleoside pentaphosphate products were hardly separable from the closely related polyphosphate byproducts. Therefore, high performance liquid chromatography (HPLC) in addition to regular ion exchange chromatography was employed to afford the desired Cp₅C (**5**) and Up₅U (**6**) in 35–38% yields.

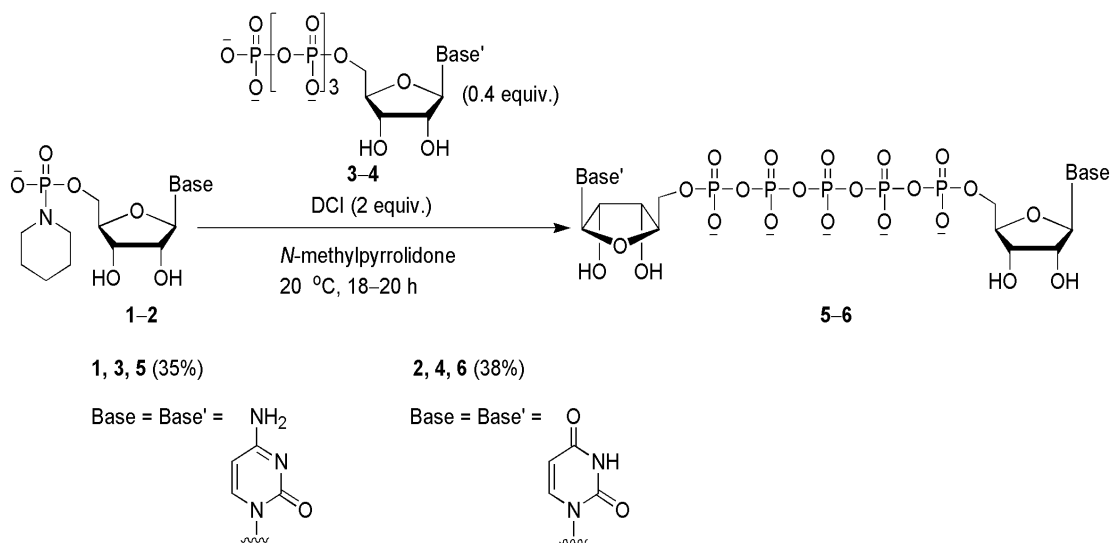


Figure 1. The P (V)–N activation method for the synthesis of dinucleoside pentaphosphates (**5–6**).

Summary

In conclusion, the work reported in this paper presents an efficient protocol for the preparation of dinucleoside pentaphosphates. Compared to the previously reported synthetic methods, the P (V)–N activation approach provided a facile and efficient approach for the synthesis of dinucleoside pentaphosphates.

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References

- [1] N. Stern, D. T. Major, H. E. Gottlieb, D. Weizman, B. Fischer, *Org. & Biomol. Chem.* 8 (2009) 4637.
- [2] C. H. V. Hoyle, R. H. Hilderman, J. J. Pintor, H. Schluter, B. F. King, *Drug Dev. Res.* 52 (2001) 260.
- [3] V. Ralevic, J. Jankowski and H. Schluter, *Br. J. Pharmacol.* 134 (2001) 1073.
- [4] A. G. Aranguez, A. Crooke, A. Peral, C. H. V. Hoyle, J. Pintor, *Prog. Retinal Eye Res.* 26 (2007) 674.
- [5] J. G. Douglass, R. I. Patel, B. R. Yerxa, S. R. Shaver, P. S. Watson, K. Bednarski, R. Plourde, C. C. Redick, K. Brubaker, A. C. Jones, J. L. Boyer, *J. Med. Chem.* 51 (2008) 1007.
- [6] K. A. Henzler-Wildman, V. Thai, M. Lei, M. Ott, M. Wolf-Watz, T. Fenn, E. Pozharski, M. A. Wilson, G. A. Petsko, M. Karplus, C. G. Hubner, D. Kern, *Nature*, 450 (2007) 838.
- [7] E. Holler, B. Holmquist, B. L. Vallee, K. Taneja and P. Zamecnik, *Biochemistry*, 22 (1983) 4924.
- [8] J. W. Pettegrew, D. W. Miles and H. Eyring, *Proc. Natl. Acad. Sci. U. S. A.* 74 (1977) 1785.
- [9] J. G. Douglass, R. I. Patel, B. R. Yerxa, S. R. Shaver, P. S. Watson, K. Bednarski, R. Plourde, C. C. Redick, K. Brubaker, A. C. Jones, J. L. Boyer, *J. Med. Chem.* 51 (2008) 1007.
- [10] J. Stepinski, M. Bretner, M. Jankowska, K. Felczak, R. Stolarski, Z. Wieczorek, A. L. Cai, R. E. Rhoads, A. Temeriusz, *Nucleosides, Nucleotides Nucleic Acids*, 14 (1995) 717.
- [11] S. R. Shaver, J. L. Rideout, W. Pendergast, J. G. Douglass, E. G. Brown, J. L. Boyer, R. I. Patel, C. C. Redick, A. C. Jones, M. Picher, B. R. Yerxa, *Purinergic Signalling* 1 (2005) 183.
- [12] W. Pendergast, B. R. Yerxa, J. G. Douglass, S. R. Shaver, R. W. Dougherty, C. C. Redick, I. F. Sims, J. L. Rideout, *Bioorg. Med. Chem. Lett.* 11 (2001) 157.
- [13] J. F. Scott, P. C. Zamecnik, *Proc. Natl. Acad. Sci. U. S. A.* 64 (1969) 1308.
- [14] W. Pendergast, B. R. Yerxa, J. G. Douglass, et al., Synthesis and P2Y receptor activity of a series of uridine dinucleoside 5'-polyphosphates, *Bioorg. Med. Chem. Lett.* 11 (2001) 157.
- [15] L. C. Davies, J. A. Stock, S. E. Barrie, et al., Dinucleotide analogues as inhibitors of thymidine kinase, thymidylate kinase, and ribonucleotide reductase, *J. Med. Chem.* 31 (1988) 1305.
- [16] P. Feldhau, T. Frohlich, R. S. Goody, *Eur. J. Biochem.* 57 (1975) 197.
- [17] Q. Sun, S.-S. Gong, J. Sun, C.-J. Wang, S. Liu, G.-D. Liu, C. Ma, *Tetrahedron Lett.* 55 (2014) 2114.