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# The Effect of Exogenous Melatonin on Kiwifruit antioxidant system under low light environment

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**Abstract.** Melatonin (N-acetyl-5-methoxytryptamine) in plants is to act as an antioxidant providing protection against environmental agents. In our study, we used different concentration of melatonin (control, 50  $\mu$ M, 100  $\mu$ M and 200  $\mu$ M) to investigate the effect of melatonin on antioxidant system under low light. The results suggested melatonin has strong antioxidant ability to effectively scavenge  $H_2O_2$  and reduce relative electrical conductivity (REC). The 100 $\mu$ M and 200  $\mu$ M group had a significant effect. Moreover, melatonin also activated peroxidase (POD), superoxide dismutase (SOD), and catalase (CAT) compared to control group. In general, 200  $\mu$ M melatonin had the mostly significant effect on protecting antioxidant system.

#### Introduction

In agricultural production, many biological or abiotic factors accelerate leaf senescence, such as low light conditions[1, 2]. However, stress-induced leaf senescence has also been associated with increased generation of reactive oxygen species (ROS) [3]. Excessive ROS will destroy the integrity of membrane structure. As a result, the relative conductivity of the leaves increased. In plants, the cells suppress the ROS accumulation by activating the enzymatic antioxidant pathway, including POD, SOD, CAT, and suppress the stress-induced senescence accordingly.

Melatonin was first detected in edible plants in 1995, and the physiological function of which in plants have been widely studied [4]. A fundamental function attributed to melatonin is to act as an antioxidant in plants. Some research suggested that melatonin regulating the redox state of cells, decreasing ROS and maintaining cell membrane stability [5].

Our study investigated the effect of different concentration of melatonin on antioxidant system by determine the  $H_2O_2$ , REC and antioxidant enzymes ability. The results will provide a crucial proof in the application of melatonin in oxidation and anti-low light in the field.

#### **Treatment and Methods**

**Treatment.** We used the healthy and uniform kiwifruit seedlings to assign two conditions for pretreatment: (i) standard water supply and low light (control), (ii) solution of 50  $\mu$ mol·L<sup>-1</sup> melatonin in water and low light (50  $\mu$ M group), (iii) solution of 100  $\mu$ mol·L<sup>-1</sup> melatonin in water and low light (10  $\mu$ M group), (iv) solution of 200  $\mu$ mol·L<sup>-1</sup> melatonin in water and low light (200  $\mu$ M group). The pretreatment was conducted for 8 days in an open experimental field. During this period, the seedlings were treated with melatonin or water every 2 day by root irrigation (20 mL per pot). After the fifth irrigation, the seedlings were put under shade with 60% sunlight (the day was



set as 0 days). The plants were sampled at day 0, 2, 4, 6, 8 between 10:00 and 11:00 h, by removing the fifth to ninth leaves upward along the stem from three trees per treatment. Every treatment used 15 pots, 3 seedlings per pot. The samples were quickly frozen after collection and stored in a cryogenic refrigerator at -80°C for subsequent index determination. All reactions were performed by using the leaf mixture of three kiwifruit seedlings with three technical and three biological replicates.

**Determination of H<sub>2</sub>O<sub>2</sub>, REC and antioxidant enzymes.** Determination of H<sub>2</sub>O<sub>2</sub> concentration was based on the method of Lin et at.[6] and the measurement of REC referred to Chen et al.[7] The activities of POD, SOD, CAT were determined using the method of Wang and Huang [8].

#### **Results**

Exogenous melatonin scavenged  $H_2O_2$  and reduced relative conductivity. When plants are under stress, cells are induced to produce and accumulate ROS. In our treatment, the control group accumulated the highest content under low light, about 368.81 μmol.g<sup>-1</sup> FW (FW: fresh weight), while the content of melatonin application groups (50 μM group, 100 μM group and 200 μM group) were significantly lower than control group. However, there was also markedly difference between each concentration. The lowest  $H_2O_2$  content was detected in 100 μM group (51.15 μmol.g<sup>-1</sup> FW), and the content of 50 μM group and 200 μM group were higher than 100 μM group (222.69 μmol.g<sup>-1</sup> and 102.71 μmol.g<sup>-1</sup> respectively). Besides, we measured the REC. The results suggested 50 μM melatonin had no difference compared to control group, both of which had higher REC (56.32% and 54.59% respectively), while the 100 μM and 200 μM group had significant effect on the reduce of REC, about 25.7% and 23.9% respectively lower than control group.

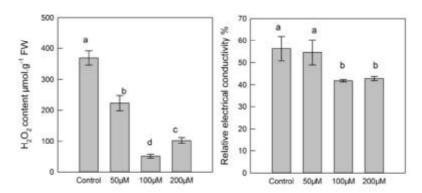


Figure 1.Effect of exogenous melatonin on  $H_2O_2$  and REC under weak light. FW means fresh weight (the same as below). Data are show as means  $\pm$  SE (n=9), different letters indicate significant differences at p < 0.05 level.

**Exogenous Melatonin improved the activity of antioxidant enzymes**. In plants, the antioxidant enzymes system alleviated the damage of peroxides to the membrane system. However, after melatonin pretreatment, the activity of all the enzymes were markedly in control group lower than melatonin pretreatment groups (50 μM group, 100 μM group and 200 μM group). Regarding POD, 200 μM group had highest activity and 91.7% higher than control group, 100 μM group followed (51.6% higher than control group), and 50 μM group had lowest activity compared with 100 μM and 200 μM group. For SOD, just the group of 100 μM and 200 μM induced the increase of sod activity, but there was no difference between 100 μM and 200 μM group. For CAT, the activity of all the groups enhanced, but the just 50 μM and 200 μM group had significant difference with control group (15.3% and 20.2% respectively higher than control group).



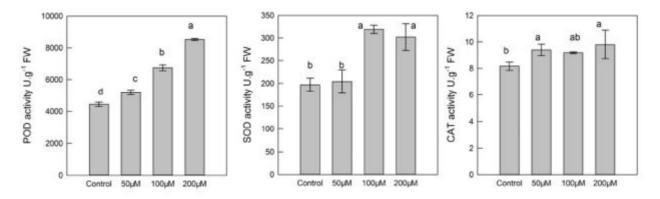


Figure 2.Effect of exogenous melatonin on antioxidant enzymes under weak light. Data are show as means  $\pm$  SE (n=9), different letters indicate significant differences at p < 0.05 level.

#### **Discussion**

Stress-induced leaf senescence will generate excessive reactive oxygen species. Excessive reactive oxygen species will accelerate the membrane lipid oxidation, and destroy the integrity of membrane structure. Thus, senescence is characterized by loss of membrane integrity, which is generally considered as increased electrolyte leakage and result the increase of REC [9]. In our study, the content of melatonin application groups was significantly lower than control group attributing to the antioxidant ability of melatonin. Besides, The 100  $\mu$ M group had the mostly significant effect on scavenge  $H_2O_2$ , and the 200  $\mu$ M group followed. Due to the lower content of  $H_2O_2$  in 100  $\mu$ M group and 200  $\mu$ M group, the REC of which were also markedly lower than control group. The beneficial effect of melatonin on scavenging hydrogen peroxide and protecting cell membrane stability also monitored in *Malus hupehensis*[10].

Moreover, we detected that melatonin also activated the antioxidant enzymes and the ability of three enzymes was enormously enhanced than control group. Maybe, it is because that melatonin induced the relative expression levels of antioxidant enzyme related genes [11]. Additionally, the 200  $\mu$ M group had the marked effect on inducing the increase of POD ability, but regarding SOD, the 100  $\mu$ M group was no difference with 200  $\mu$ M group, and the three groups also have no difference in CAT. Due to the higher enzymes activity, the lower concentration of  $H_2O_2$  and REC also can be explained.

#### **Conclusions**

In our study, melatonin significantly activated the enzymes POD, SOD and CAT and acted as an efficient antioxidant,  $H_2O_2$  was efficiently scavenged, thus, membrane integrity was maintained. After comprehensive analysis of all results, we suggest  $200 \, \mu M$  melatonin had the mostly significant effect on protecting antioxidant system. It is hoped that our results will lay a foundation for the application of melatonin.

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